

Zooplankton is the perfect candidate for validating eDNA metabarcoding for analyzing North Sea marine fauna

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S04 (Day 2)

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Environmental DNA

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Capturing drifting species and molecules—Lessons learned from integrated approaches to assess marine metazoan diversity in highly dynamic waters

Alica Ohnesorge 🔀, Uwe John, Sarah Taudien, Stefan Neuhaus, Lucie Kuczynski, Silke Laakmann 🔀

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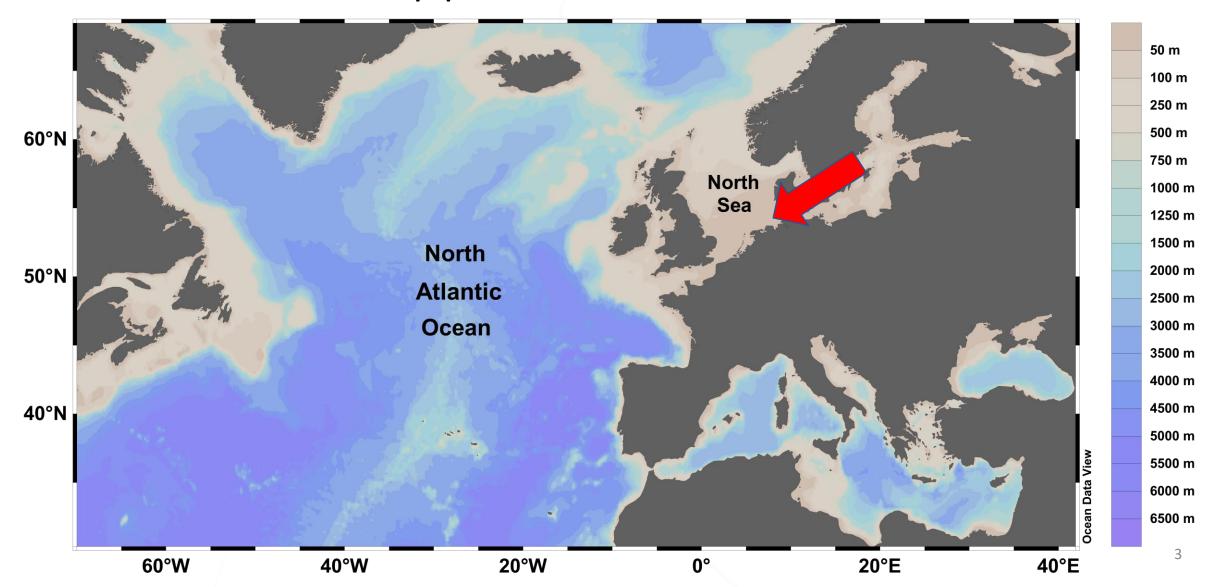
AIM:

Do we have sufficient reference data & appropriate methods to identify marine fauna in the North Sea?



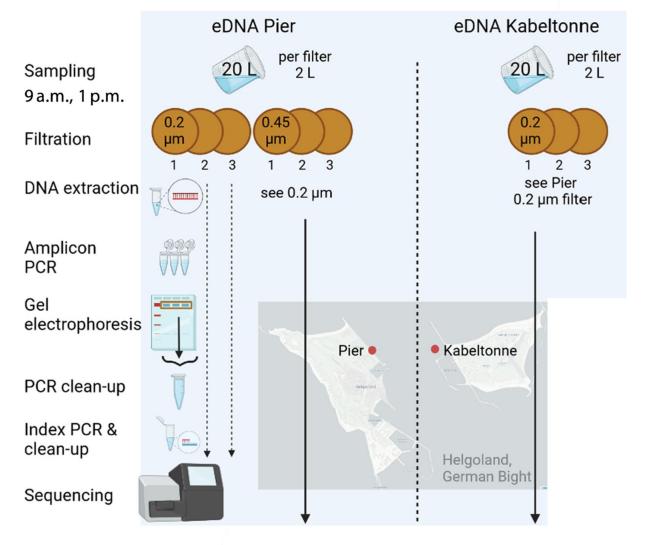


The Approach





The Approach



Sampling

Helgoland (North Sea, German Bight) 19-27 June 2019

eDNA: ca. 260 L

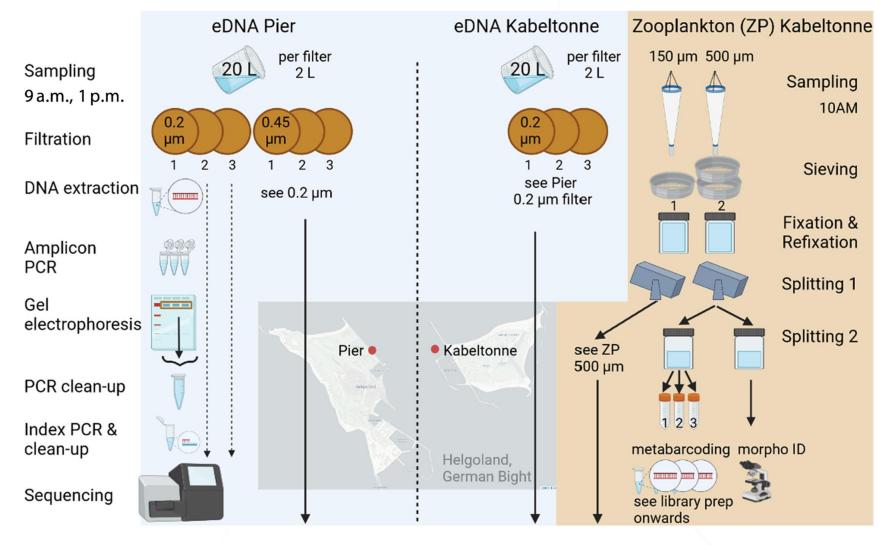
Metabarcoding:

a) COI: MetaZooGene db

o) 18S V4: SILVA & PR2



The Approach



Sampling

Helgoland (North Sea, German Bight) 19-27 June 2019

eDNA: ca. 260 L

ZP net: ca. 120,000 L (150 & 500 μm)

Metabarcoding:

a) COI: MetaZooGene db

b) 18S V4: SILVA & PR2



Validation of metabarcoding

ZP net catches: metabarcoding vs. taxa identified by morphological characteristics:

Which taxa do we **miss** by metabarcoding?

- Phoronida
- Asteroidea ⊀



- (Chaetognatha)

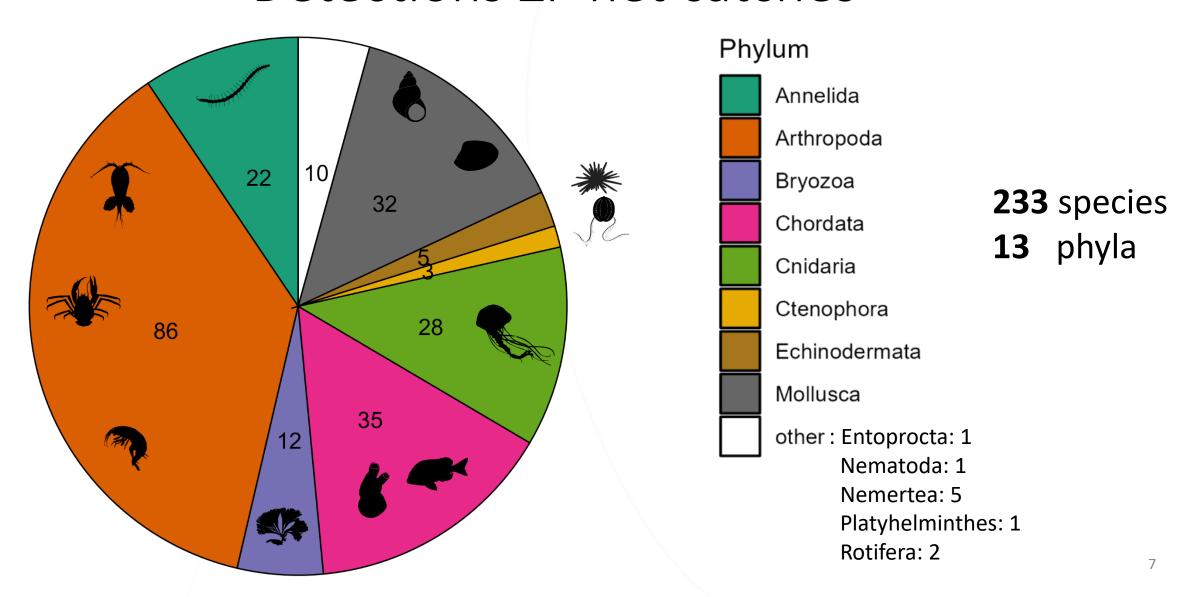
Which phyla are **additionally** recovered by metabarcoding?

- + Rotifera
- + Nemertea



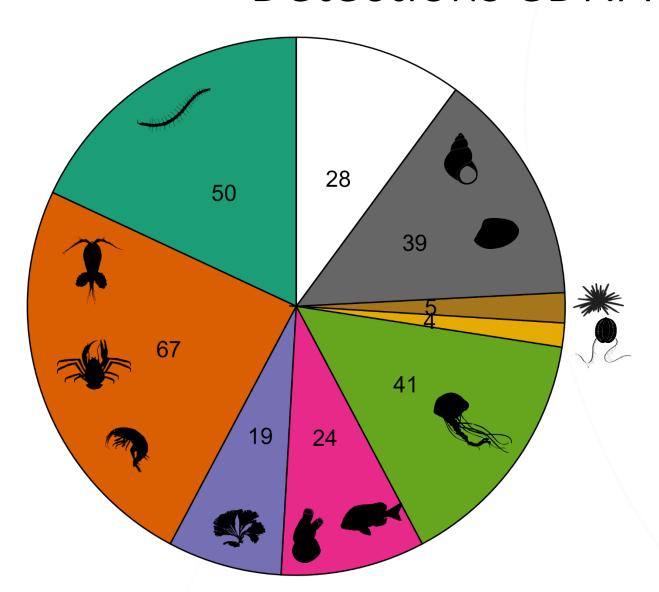


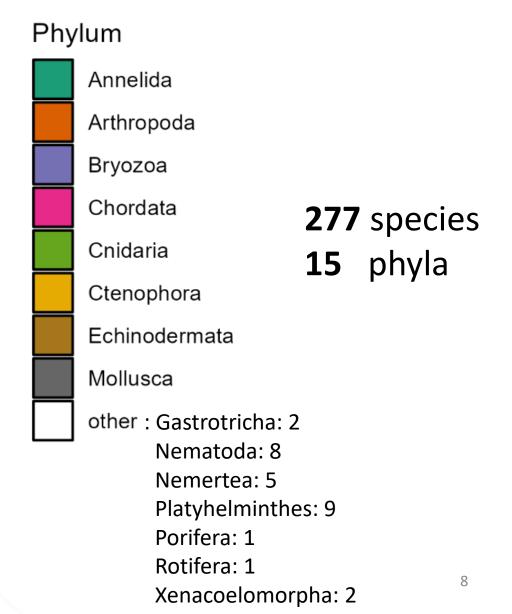
Detections ZP net catches





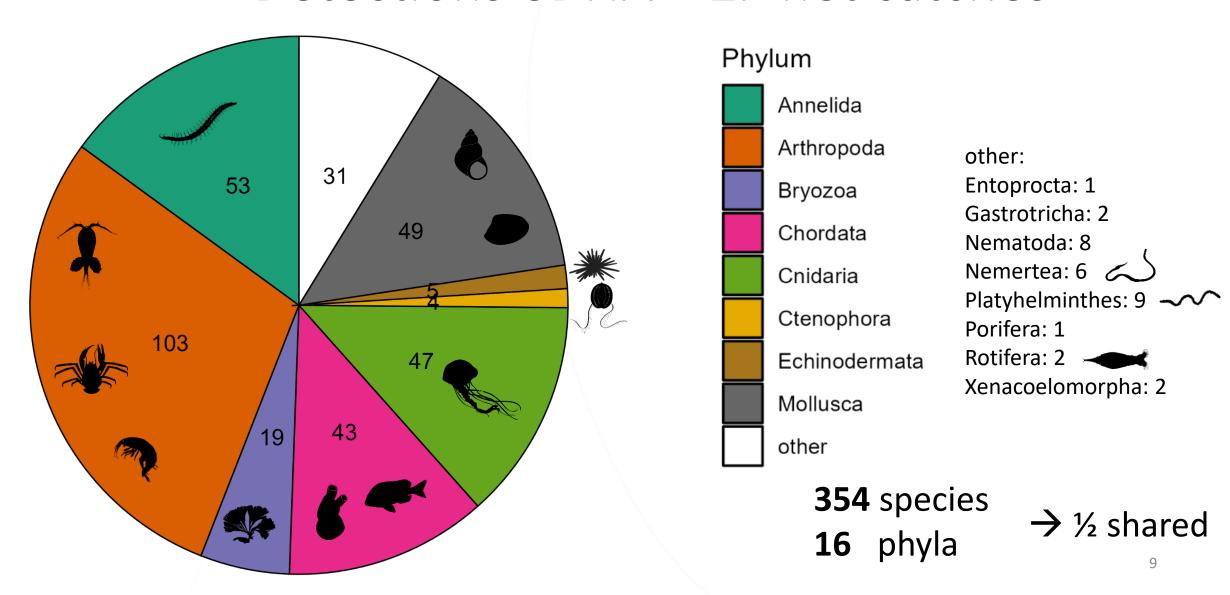
Detections eDNA







Detections eDNA + ZP net catches





Validation of metabarcoding

Are the detected species by metabarcoding known from the North Sea?

	total	Very likely	Likely	Unlikely
COI	269	238 (88.5 %)	20 (7.4 %)	11 (4.1 %)



Validation of metabarcoding

Are the detected species by metabarcoding known from the North Sea?

	total	Very likely	Likely	Unlikely
COI	269	238 (88.5 %)	20 (7.4 %)	11 (4.1 %)
18S V4	127	84 (63.0%)	20 (15.7 %)	23 (18.1 %)



congeneric species



Potentially new in German Seas?

Acartia (Acartiura) hudsonica

North Sea: this study: eDNA & ZP, COI & 18S

Baltic Sea: study submitted





Re-discovered species

Dicoryne conferta Hydrozoa before 1950



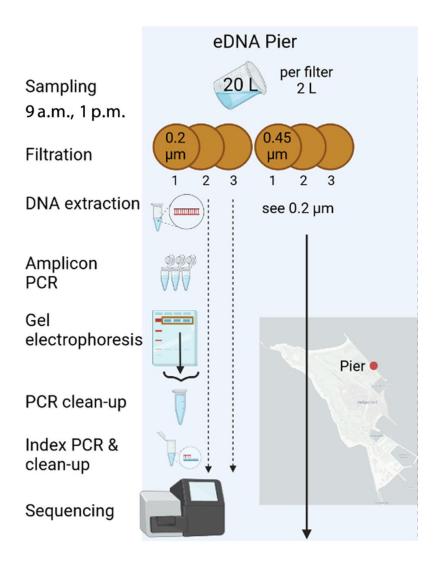
Sagartia (Cylista) viduata Anthozoa before 1950



Megadrilus purpureus Polychaeta before 1900



HIFMB Do we need replicates?



2x / day over 9 days \rightarrow 18 sampling events

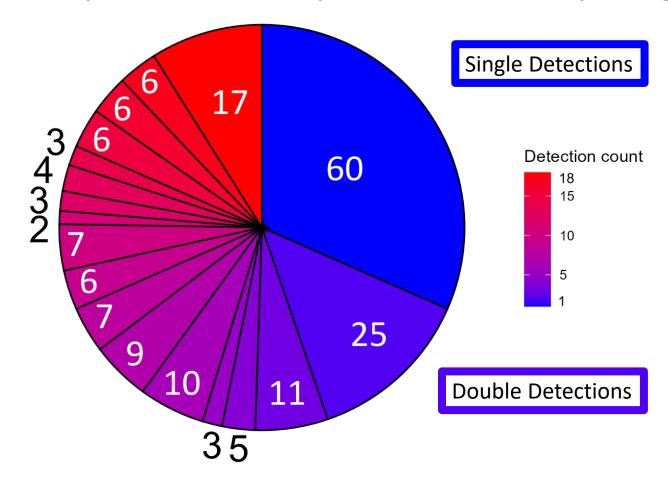
6 replicates à 2 L/ sampling event

→ 108 filter replicates in total

(following results based on COI only)



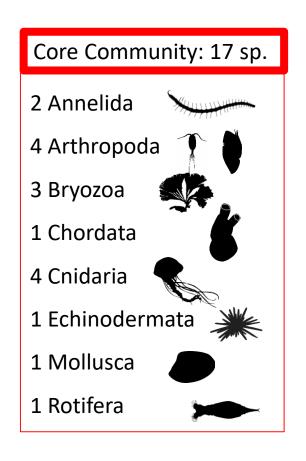
Do we need replicates: repetitive sampling

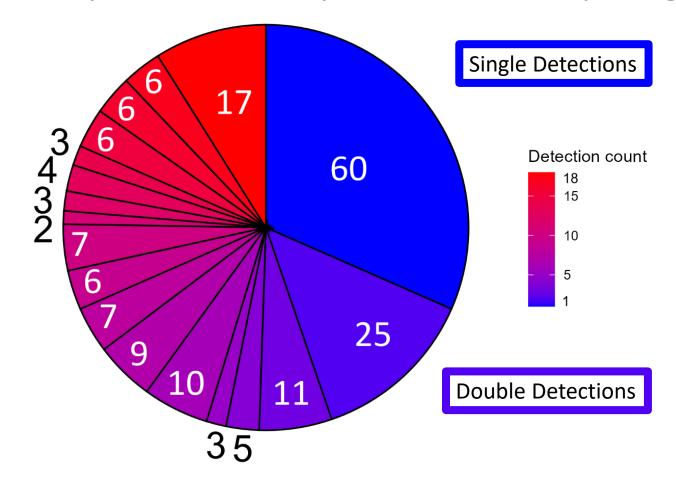


[✓] Almost half of the species (44.7 %) was detected only during one or two sampling events



Do we need replicates: repetitive sampling





- ✓ Almost half of the species (44.7 %) was detected only during one or two sampling events
- √ 17 species ('Core Community') were always detected (ca. 9 %)



Summary

- YES We are ready to use molecular identification methods to monitor North Sea zooplankton and marine fauna
- eDNA and zooplankton net catches identify slightly different communities:
 - eDNA
 - ++ benthic-associated taxa (i.e. polychaetes)
 - ZP net catches
 - ++ Arthropoda (i.e. holo- and meroplankton)
 - ++ identification of reproductive state based on mero- and ichthyoplankton
- Core community identified by low replication;
 comprehensive community identified by high replication



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