

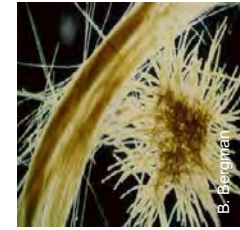
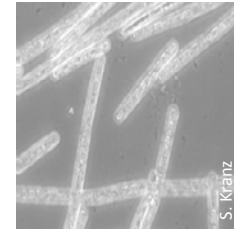


Trichodesmium under ocean acidification - A close-up on colony microenvironments -

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and Helle Ploug

Trichodesmium Background

- **Filamentous, colony-forming** cyanobacteria
- Forms **extensive blooms** in subtropical and tropical, oligotrophic oceans
- Important **N₂ fixer**
- Diverse range of **associated organisms**
- Various **toxic effects** reported (e.g. ciguatoxin-related toxins)

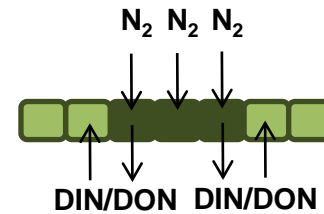
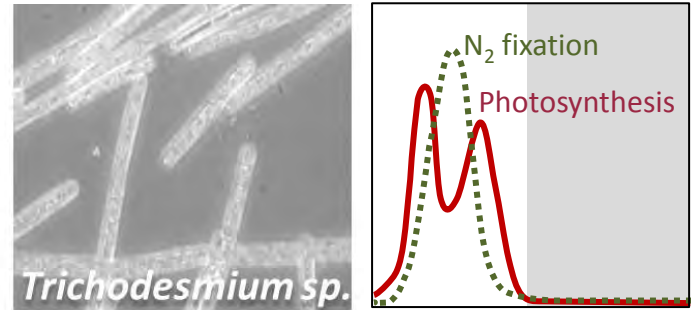


Trichodesmium

Cellular N- and C-fluxes

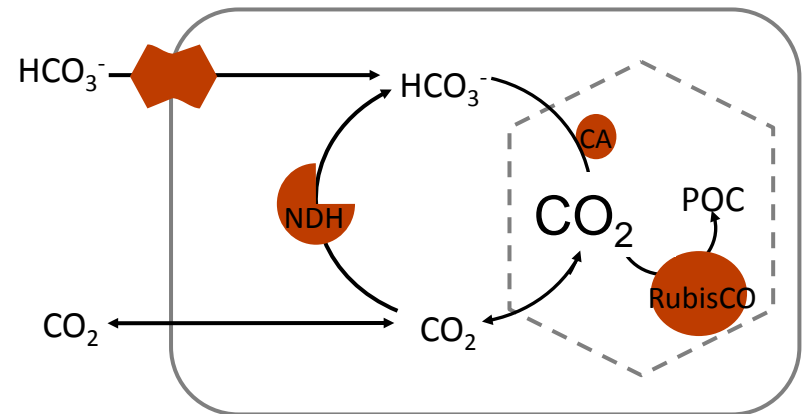
- N_2 fixation has a high energy demand & is impeded by O_2

→ Separation of N_2 fixation and photosynthesis in time (diurnal cycle) and space (diazocytes)



- RubisCO has a low CO_2 affinity & is competitively inhibited by O_2

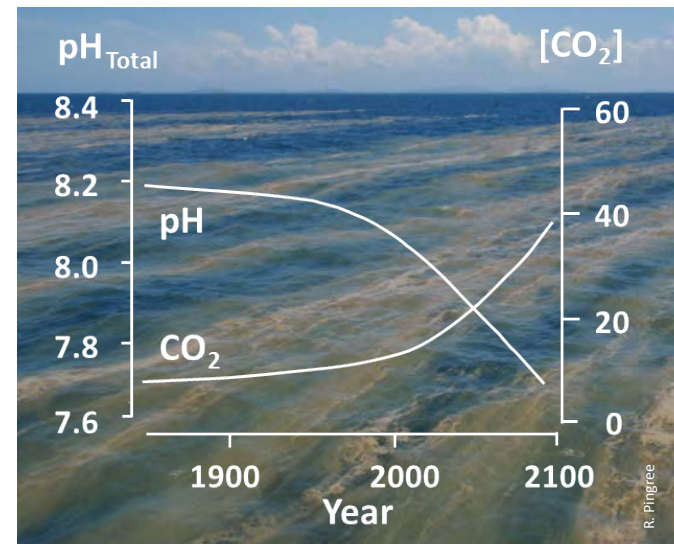
→ Operation of Carbon Concentrating Mechanisms (CCM)



Trichodesmium and OA

Previous results

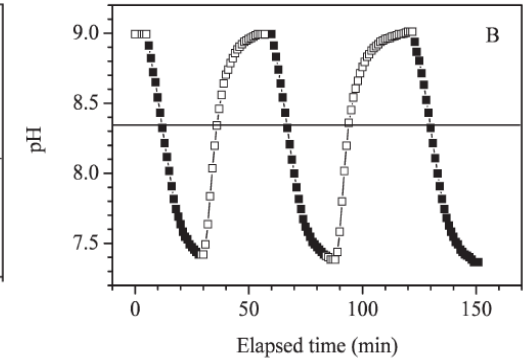
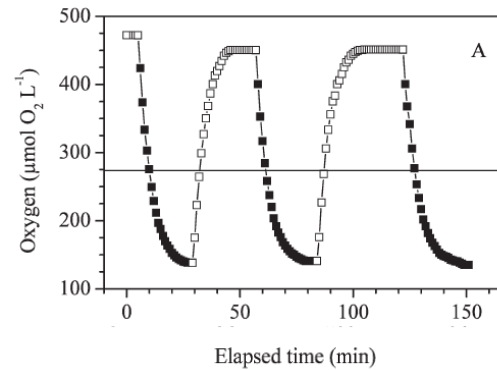
- In **lab** studies *Trichodesmium* responded **sensitively to OA**, increasing growth, N₂ fixation and POC & PON production (e.g. Hutchins et al. 2007, Kranz et al. 2009)
- **Energy reallocation** from the **CCM to N₂ fixation** suggested as a key driver behind OA responses (e.g. Kranz et al. 2011, Eichner et al. 2014)
- **Field** experiments with N₂-fixing communities found **no OA effects** (e.g. Böttjer et al. 2014, Gradoville et al. 2014)



Microenvironments of colonies

Previous results

- Conditions in colonies can differ strongly from the bulk with respect to e.g. $[O_2]$ & pH (e.g. Paerl & Bebout 1988, Ploug 2008)

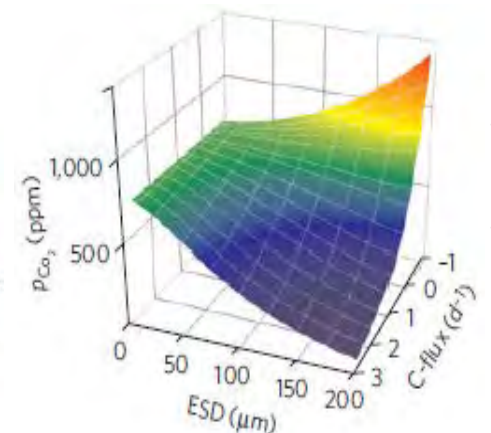
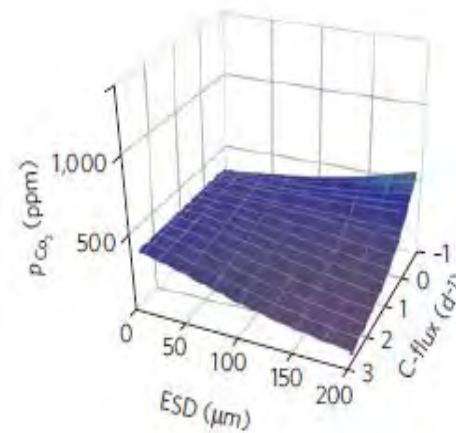


Ploug 2008

- Variability in $p\text{CO}_2$ depends on size and bulk $p\text{CO}_2$ (e.g. Flynn et al. 2012)

current

future



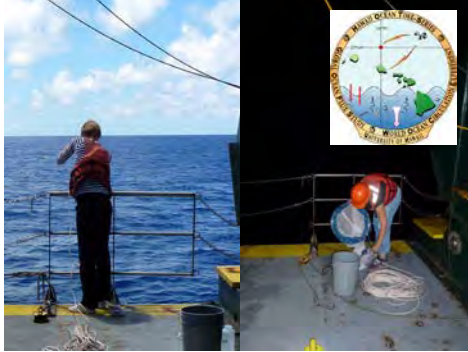
Flynn et al. 2012

→ *Requirements for the CCM differ between colonies and single filaments*



Microenvironments under OA

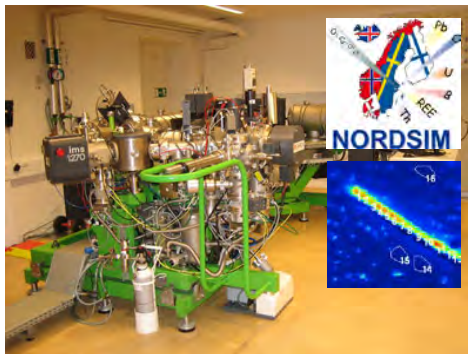
Methods



- *Trichodesmium* colonies
500 vs. 1000 $\mu\text{atm } p\text{CO}_2$



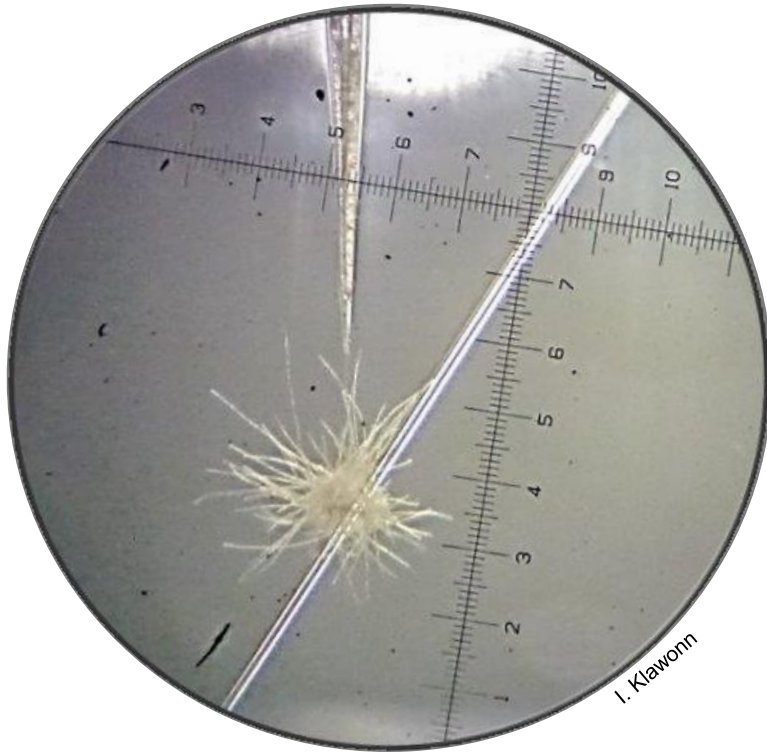
- Microsensor measurements:
 O_2 & pH



- Stable isotope incubations:
 $^{15}\text{N}_2$ & ^{13}C ; SIMS analysis

Microenvironments under OA

Colony characteristics

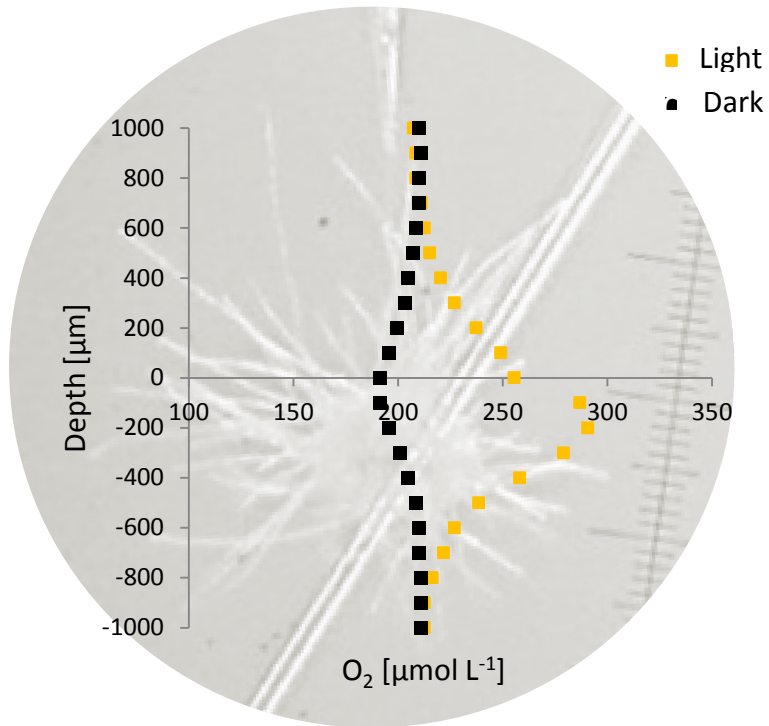


- Diameter from 0.4 to 1 mm
- 400 to 15,000 cells colony⁻¹
- Different types of filaments (70% á ~10 µm, 30% á ~20 µm width)



Microenvironments under OA

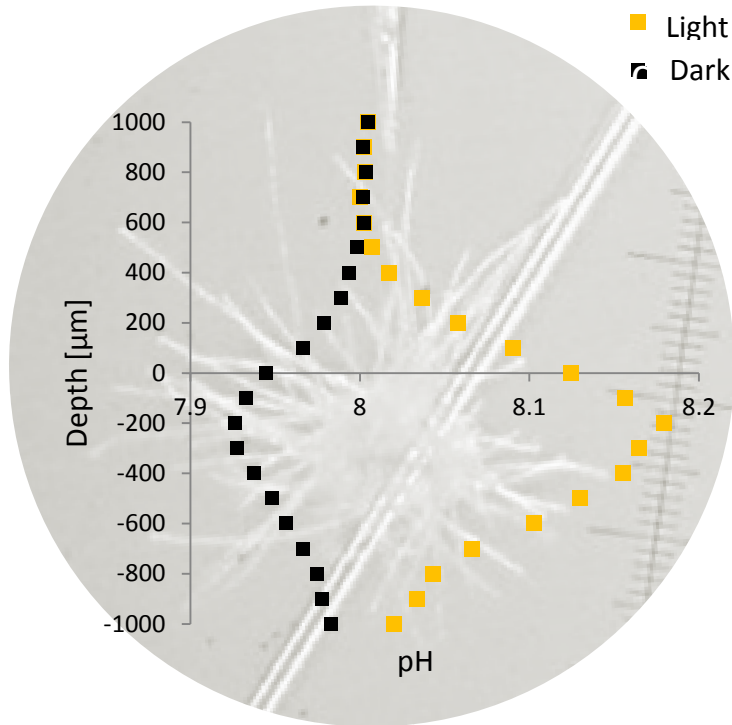
O₂ profiles



- [O₂] in the center of colonies
 - Light: 146 ± 35 % air saturation
 - Dark: 78 ± 8 % air saturation
- Total range 61 % to 203 % air sat.
- Diurnal variation in photosynthesis
- No CO₂ effects

Microenvironments under OA

pH profiles



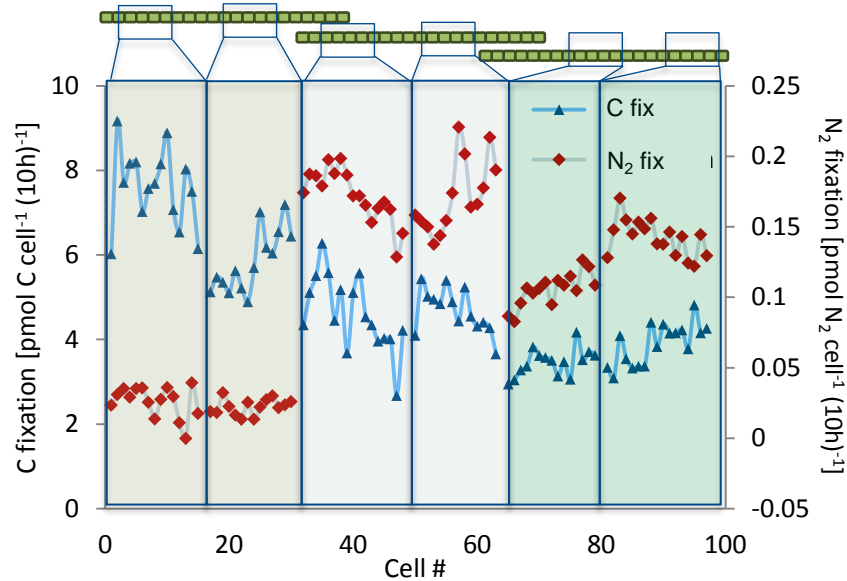
- pH in the center of colonies

	@ 8.1 bulk	@ 7.8 bulk
<u>Light</u> :	8.3 ± 0.5	7.8 ± 0.5
<u>Dark</u> :	7.8 ± 0.2	7.4 ± 0.3

- Total range 7.1 to 8.6
- Larger variability in $[H^+]$ at high pCO_2
 - @ 8.1 bulk: $\Delta[H^+] = 12 \text{ nmol L}^{-1}$
 - @ 7.8 bulk: $\Delta[H^+] = 20 \text{ nmol L}^{-1}$

Microenvironments under OA

Single cell C- and N-fluxes

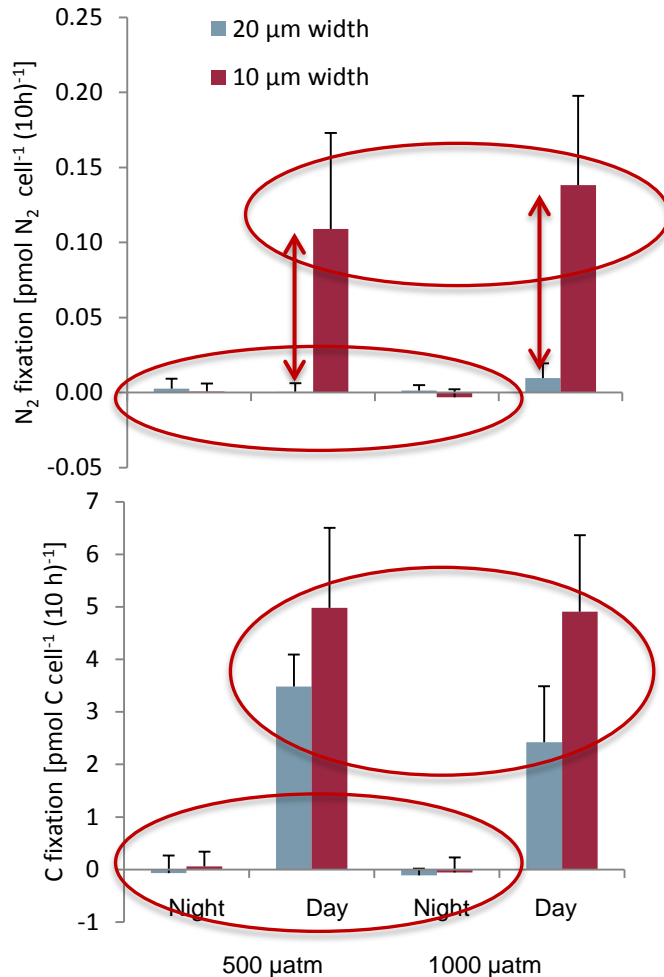


- High variability between filaments but not between cells within filaments



Microenvironments under OA

Single cell C- and N-fluxes



- High C- and N₂-fixation rates during the day
- 'Wide filaments' are non-diazotrophs
- No CO₂ effects



Microenvironments under OA

Conclusions & Implications

Conclusions

- Highly variable $[O_2]$ and pH
 - Increasing variability in $[H^+]$ with pCO_2
- Demand for high flexibility in pH-dependent processes, incl. CCMs

- High N_2 fixation + high $[O_2]$
- Demand for efficient protection/repair of nitrogenase
- High C fixation + high pH
- Demand for highly active CCM

Variability

Energy demand

To be continued...

- Low OA sensitivity due to adaptation to high variability?
- Can flexibility be increased to match future pH-variability?

- Do high energy demands enforce positive OA responses (mediated by energy re-allocation)?



Thank you for your attention

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