

Generation of ROS (reactive oxygen species) by *Chattonella marina* as a possible factor responsible for the fish-killing mechanism

**Division of Biochemistry,
Faculty of Fisheries,
Nagasaki University, Japan
Tatsuya Oda**

In Japan, HABs due to *C. marina* and *H. akashiwo* often cause mass mortality of aquacultured fish, especially yellowtail



Recent *Chattonella* red tide

Chattonella is known to be highly toxic to fish, especially yellowtail. Recent *Chattonella* red tide seems to be involved in the mass mortality of shellfish

Proposed toxic factors of *Chattonella*

**I. Toxins: hemolytic toxin,
polyunsaturated fatty acids,
neurotoxins (brevetoxins)**

II. Mucus substances

III. Reactive oxygen species (ROS)

IV. Still unknown toxins

**Ruptured *Chattonella* cell suspension has
no toxic effects**

Major Questions

- (1) Is *C. marina* really producing ROS?**
- (2) How about other raphidophytes?**
- (3) Why is *C. marina* producing ROS?**
- (4) Is ROS involved in fish-killing by *C. marina*?**
- (5) What is the mechanism of ROS production in *C. marina*?**
- (6) What is the ROS-mediated toxic mechanism of *C. marina*?**
- (7) Are there any other possible toxic factors?**

**Is *C. marina* really
producing ROS?**

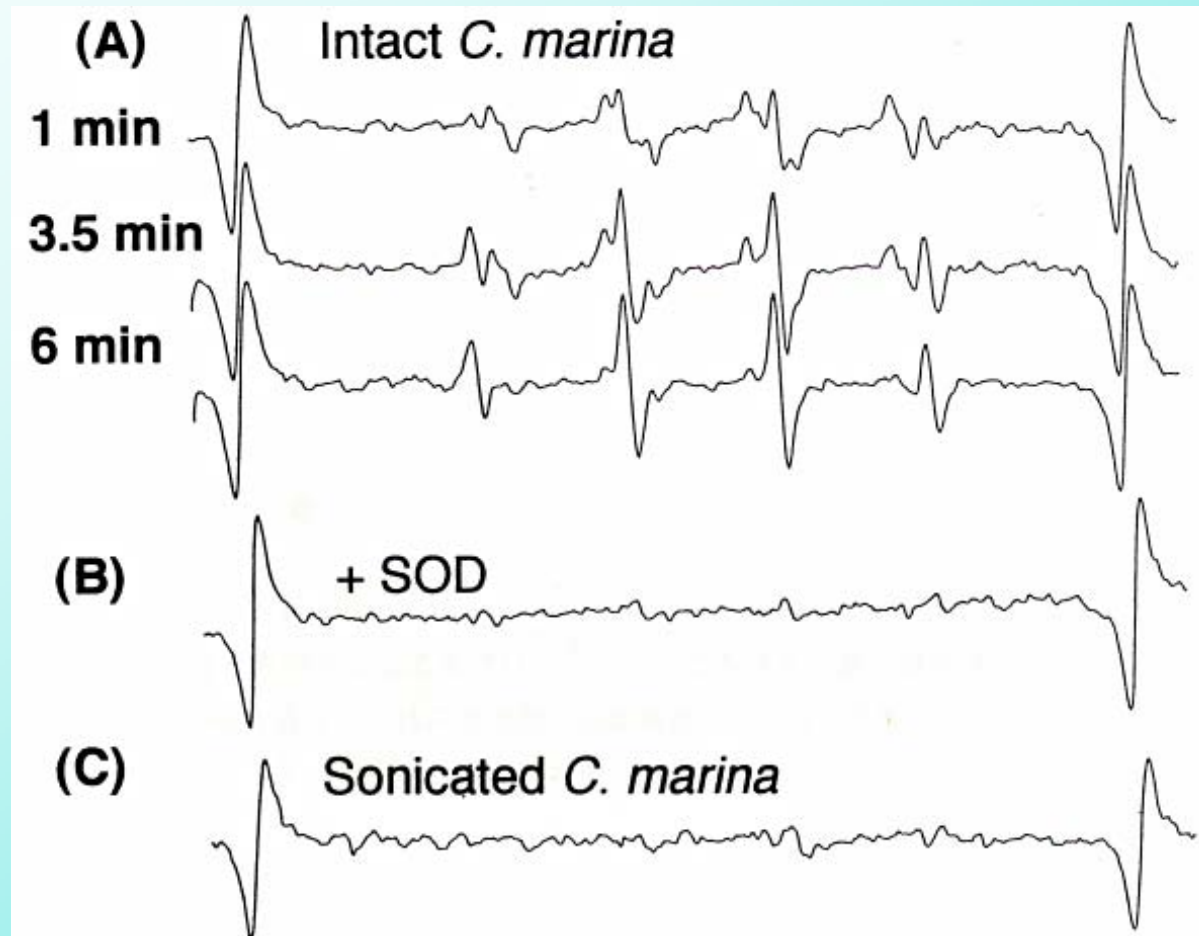
Detection of ROS in *C. marina* cell suspension by various assay methods

Cytochrome c reduction assay: +

Chemiluminescence assay: +

**ESR spectroscopic assay (the most
reliable method): +**

Detection of ROS in *C. marina* cell suspension by ESR spectroscopy



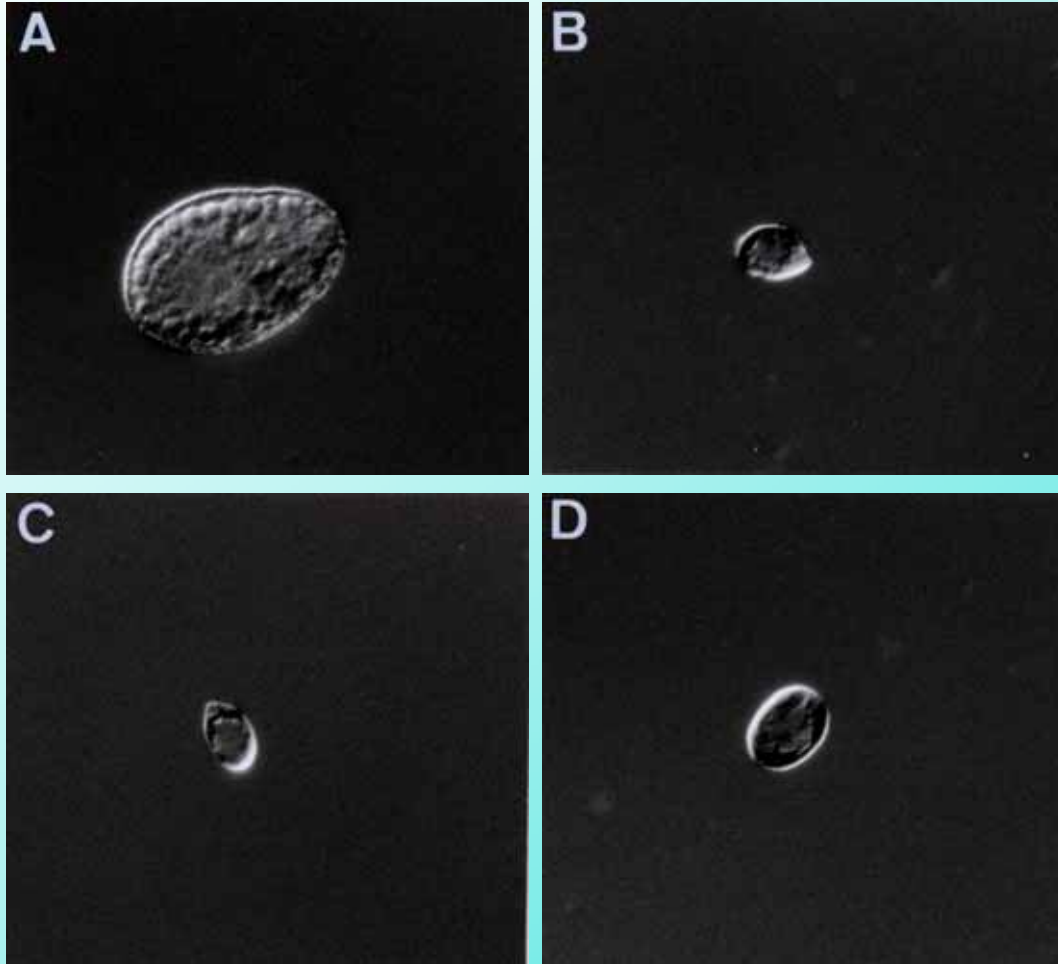
***C. marina* continuously produces ROS (mainly superoxide)**

Ruptured cell cannot produce ROS anymore, meaning that intact live condition is essential for the effective ROS generation

**How about other
raphidophytes?**

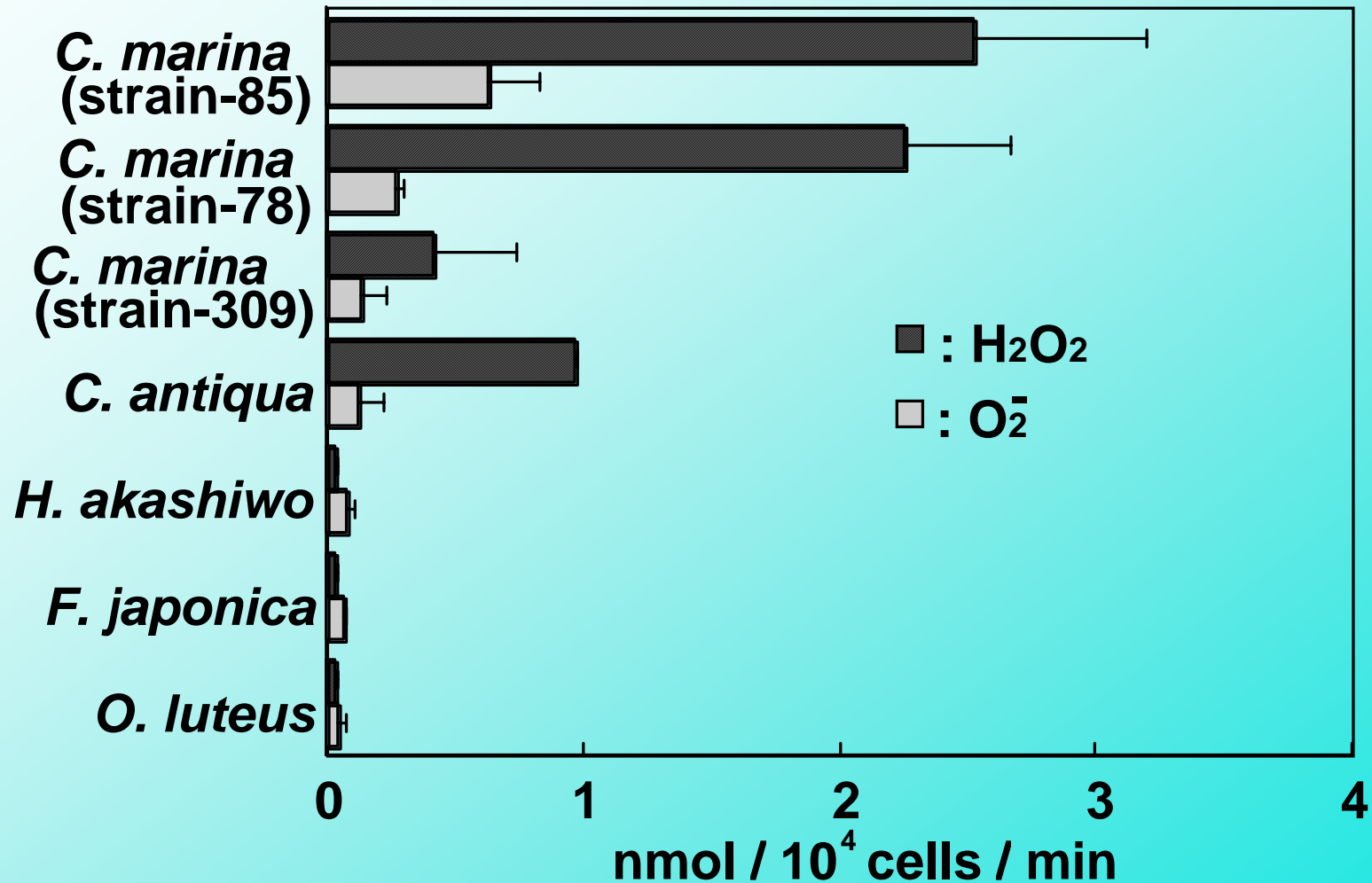
**Is ROS production a
common feature among
raphidophytes?**

**(A) *C. marina*, (B) *O. luteus*, (C) *H. akashiwo*,
and (D) *F. japonica***

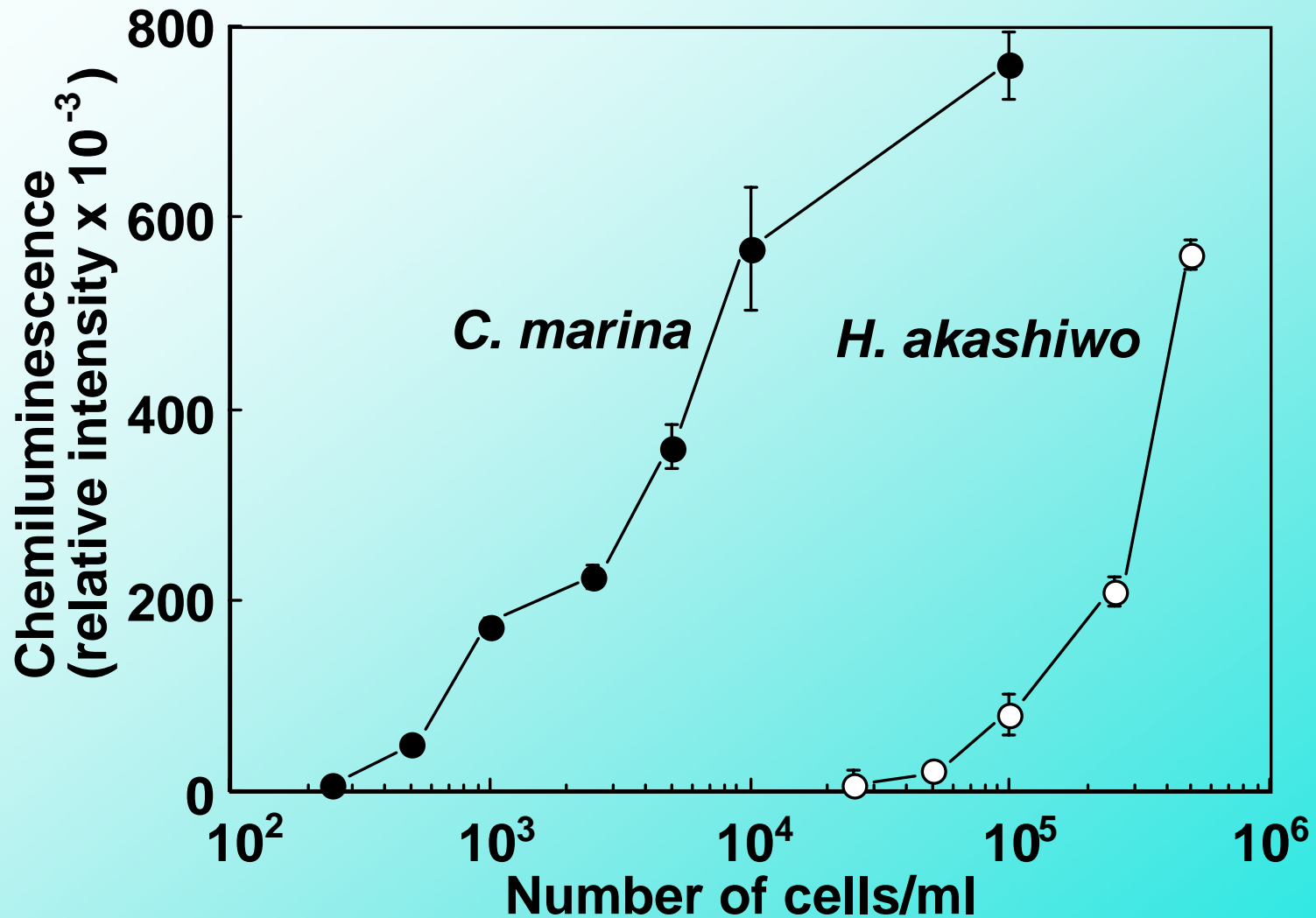


—
20 μm

ROS generation by raphidophycean flagellates



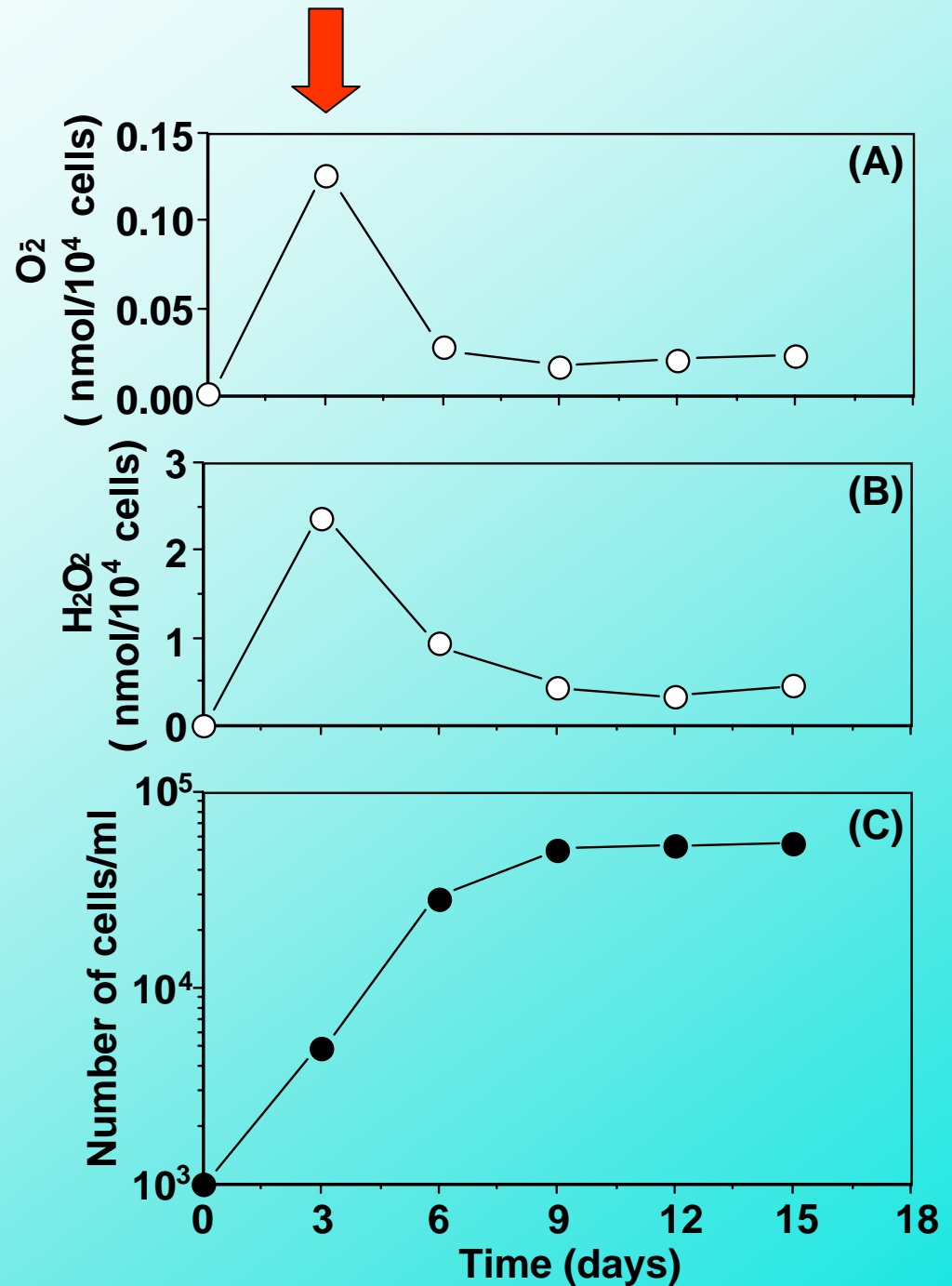
Cell density dependent ROS generation by *C. marina* and *H. akashiwo*



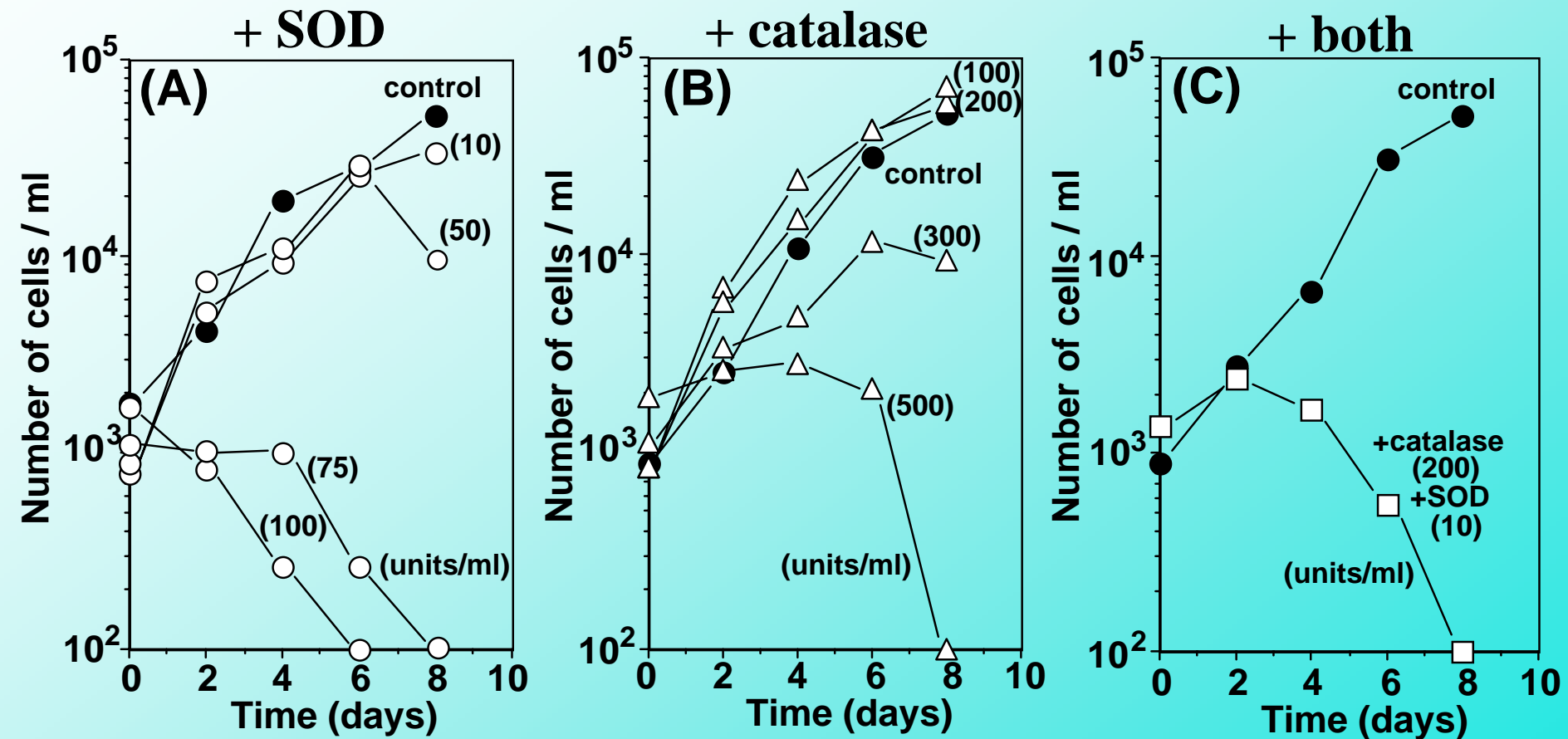
ROS levels in raphidophytes are quite different, even in *C. marina*, different ROS levels are observed among the strains

**Why is *C. marina*
producing ROS?**

The generation of ROS became the highest during the exponential growth phase



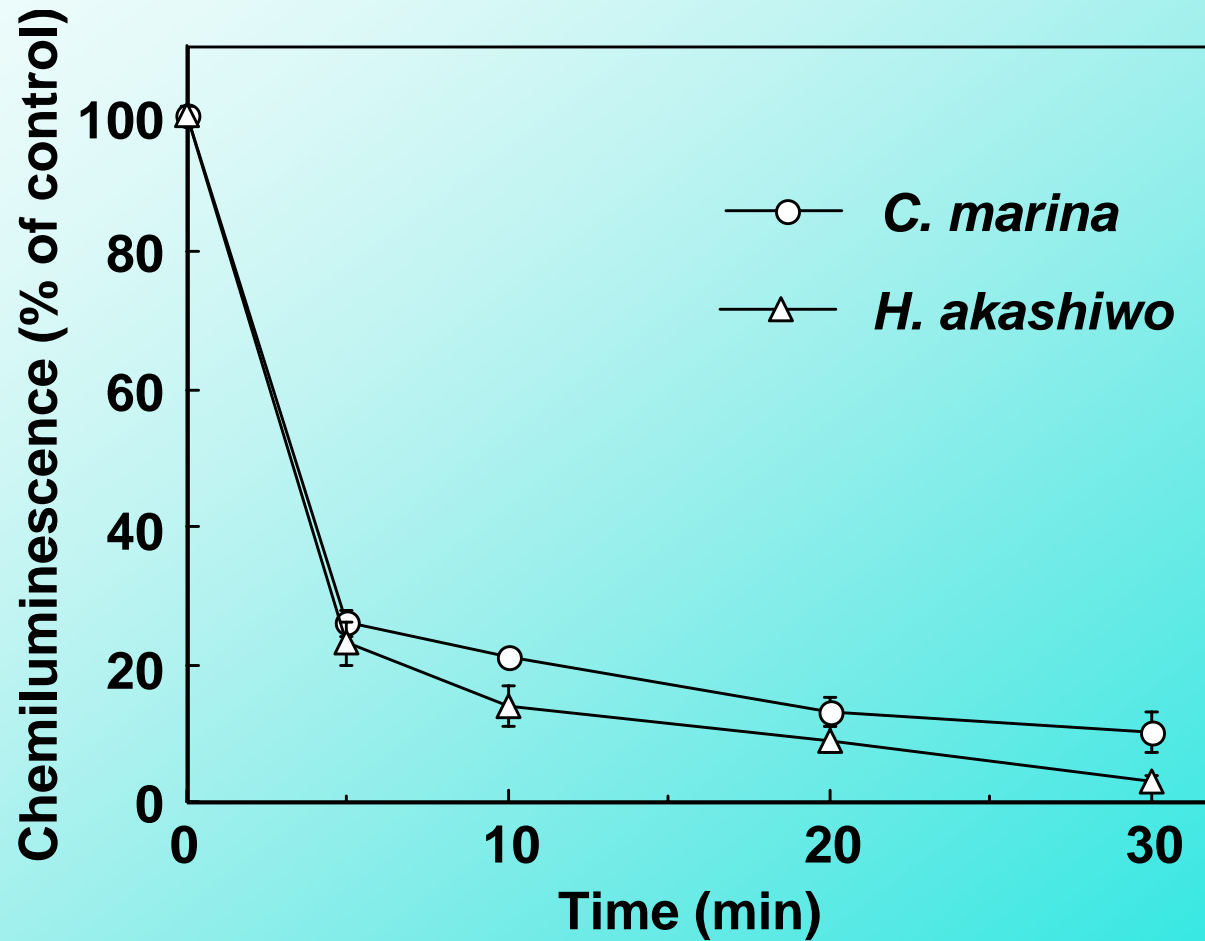
The growth of *C. marina* was inhibited by SOD and catalase



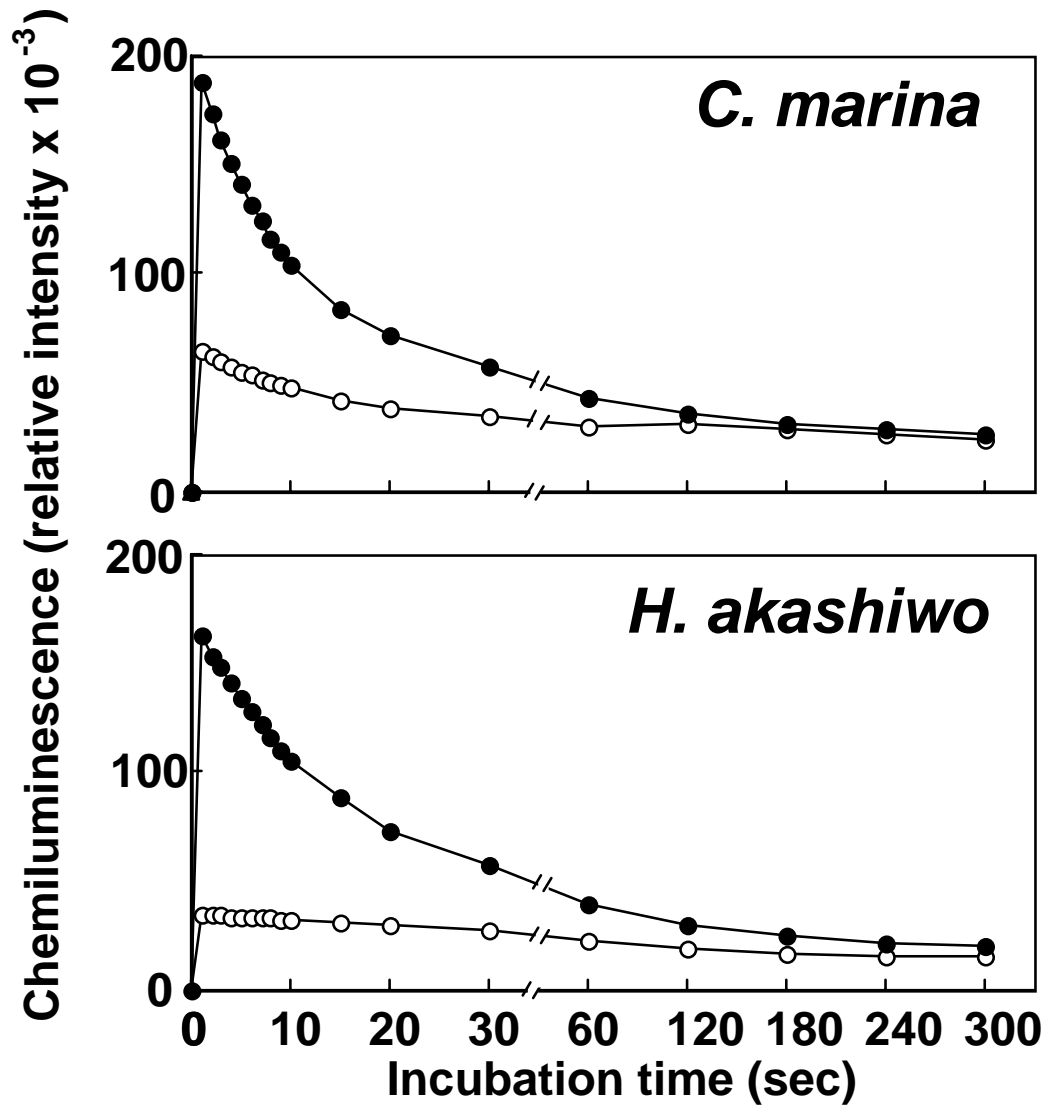
ROS generation by *C. marina* is deeply linked with the metabolic potential and may play an essential role in the own survival rather than defense purpose

**What is the mechanism
of ROS production in
C. marina?**

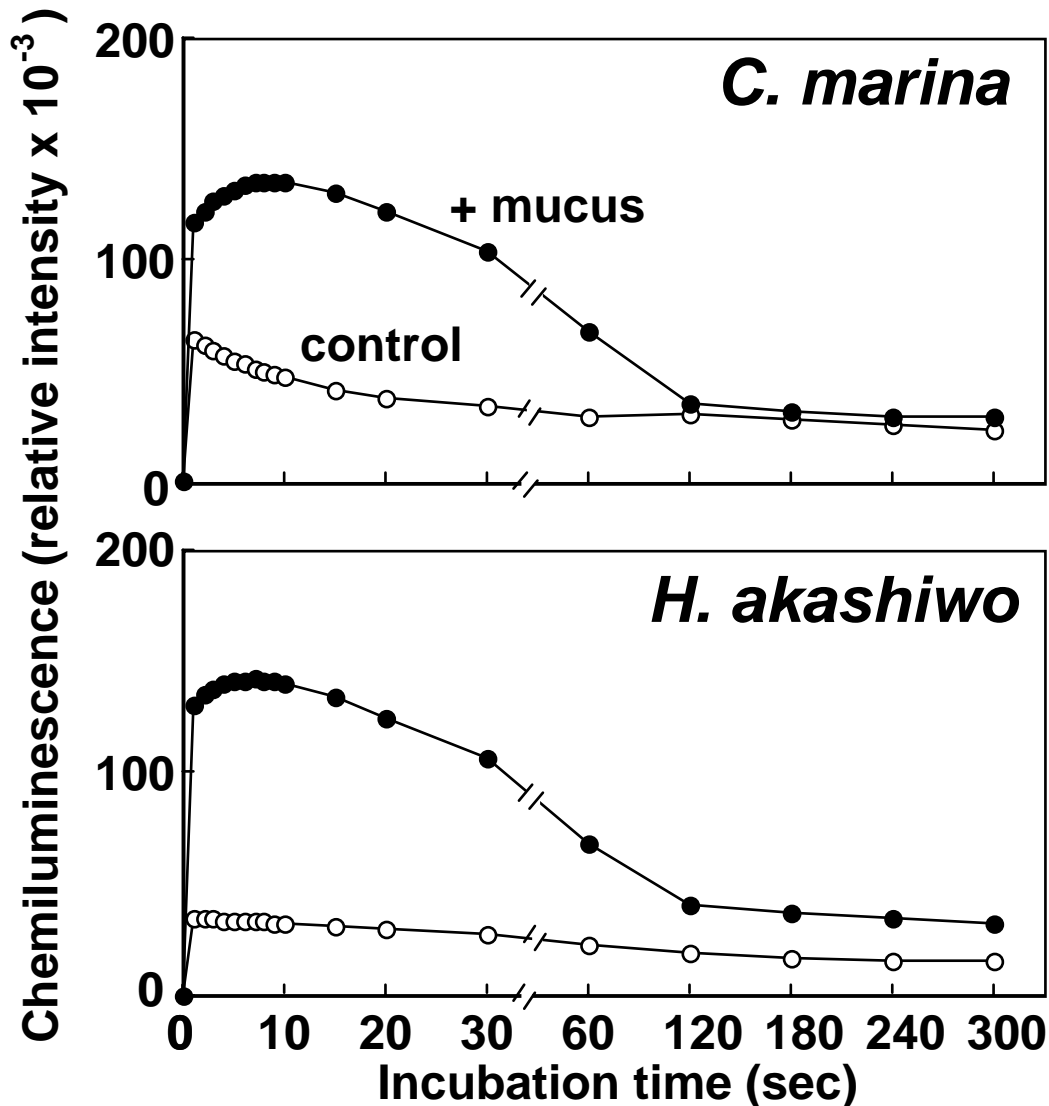
Protease (proteinase K) inhibits ROS generation by *C. marina* and *H. akashiwo*



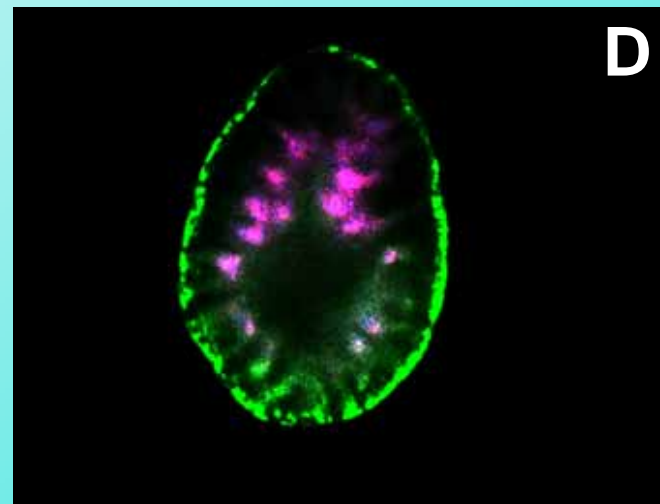
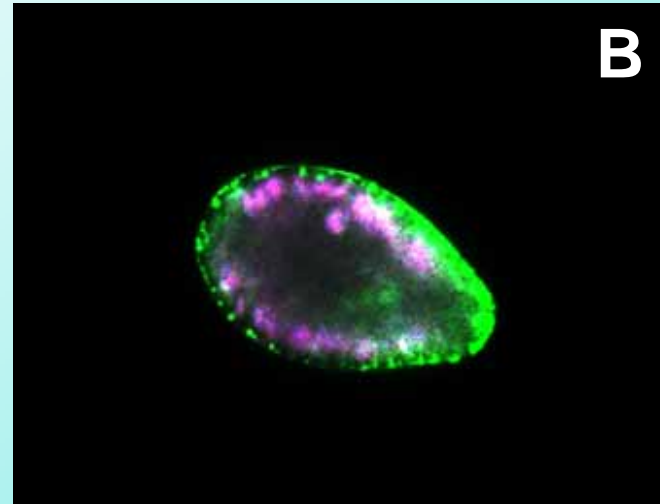
Lectin (Con A) stimulates *C. marina* and *H. akashiwo* to induce increased ROS generation



Gill mucus from yellowtail stimulates *C. marina* and *H. akashiwo* to produce increased levels of ROS

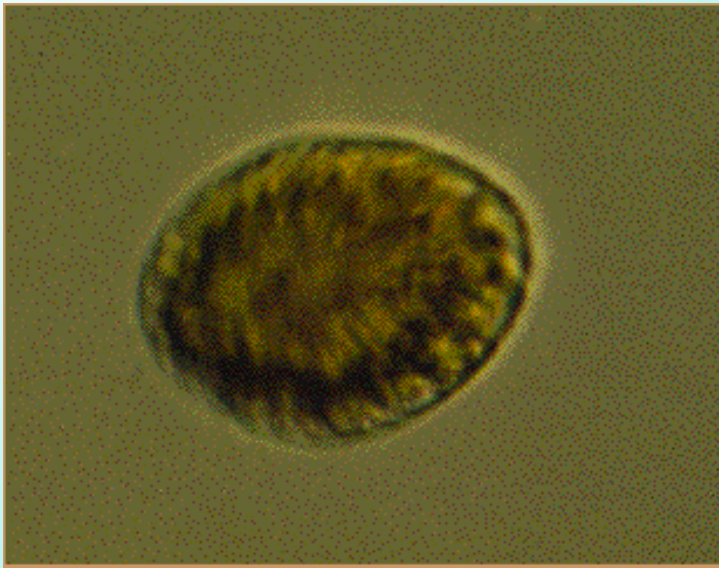


Fluorescence microscopic observation of *C. marina* (A, B) and *C. ovata* (C, D) after incubation with MCLA as a specific fluorescent probe for superoxide

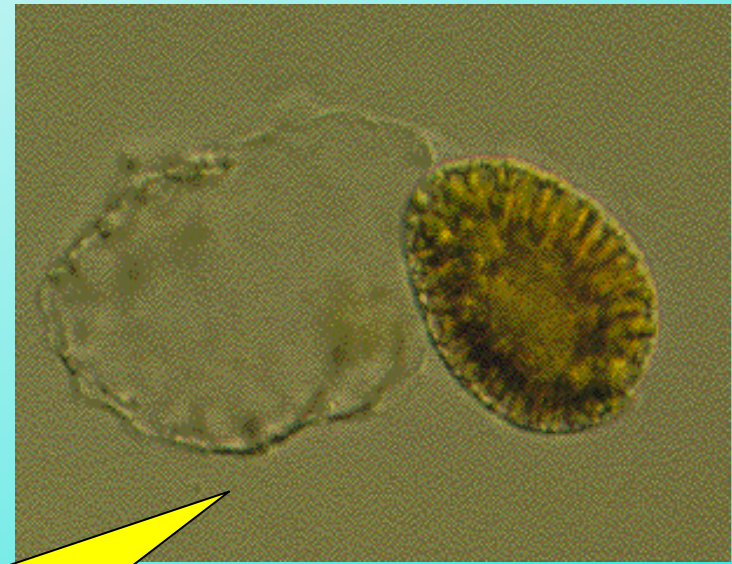


Cell surface structure of *Chattonella*: Glycocalyx

Normal condition

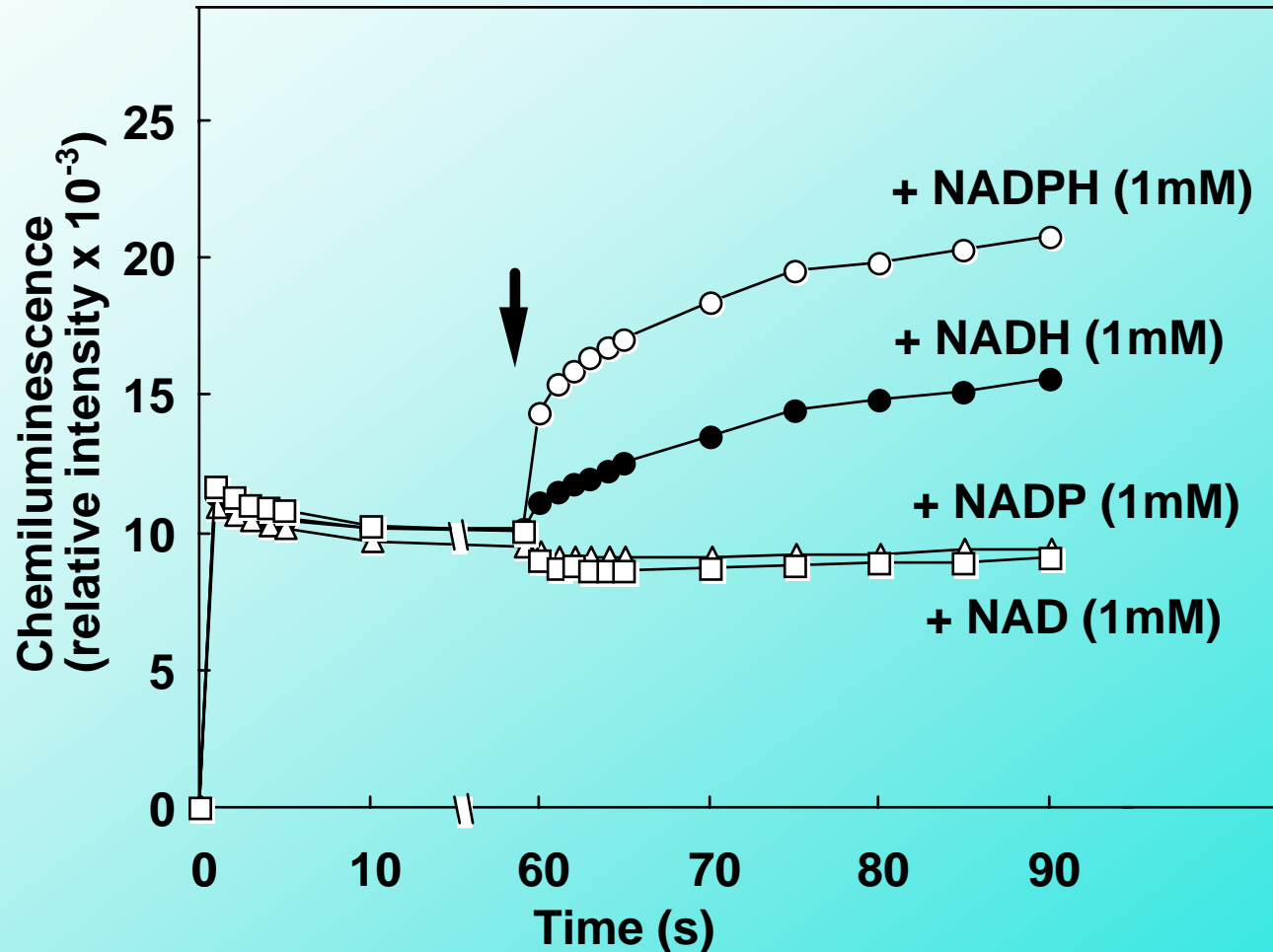


Physical stimulation:
vigorous agitation or
ultrasonic



Discharged glycocalyx

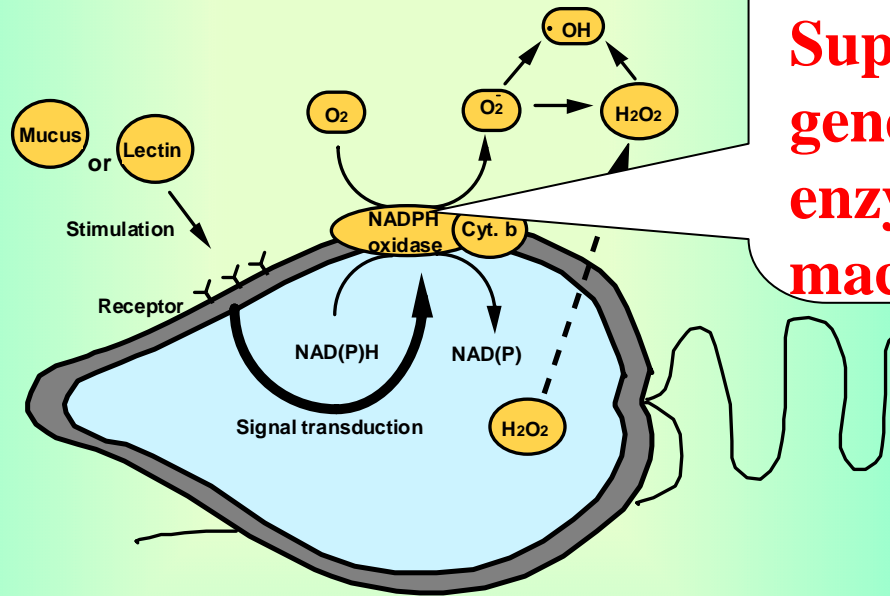
Presence of NADPH-dependent ROS generation system in Glycocalyx



Enzymatic system located on the cell surface of *C. marina* is responsible for ROS generation, and the activity can be increased by the stimulation of lectin or gill tissue mucus

Superoxide generating enzymatic system on *Chattonella* cell surface

Generation systems of reactive oxygen species (ROS) in *C. marina*



**Similar to
Superoxide
generating
enzyme in
macrophage**

**What is the ROS-
mediated toxic
mechanism of *C. marina*?**

Glycocalyx, cell surface structure of *C. marina*

Location of ROS generation system in *C. marina*

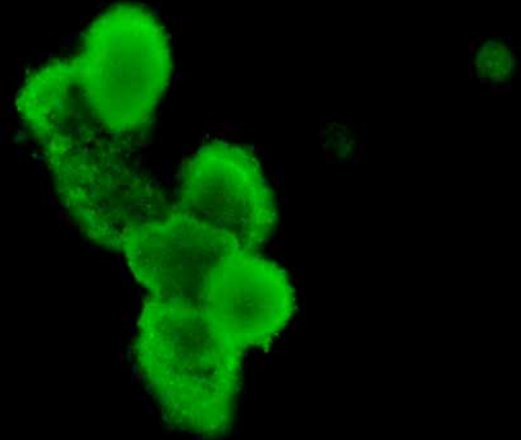
Involvement of glycocalyx in the fish-killing mechanism

**Glycocalyx can easily
discharged from *C. marina*
under the physical or chemical
stimulations**

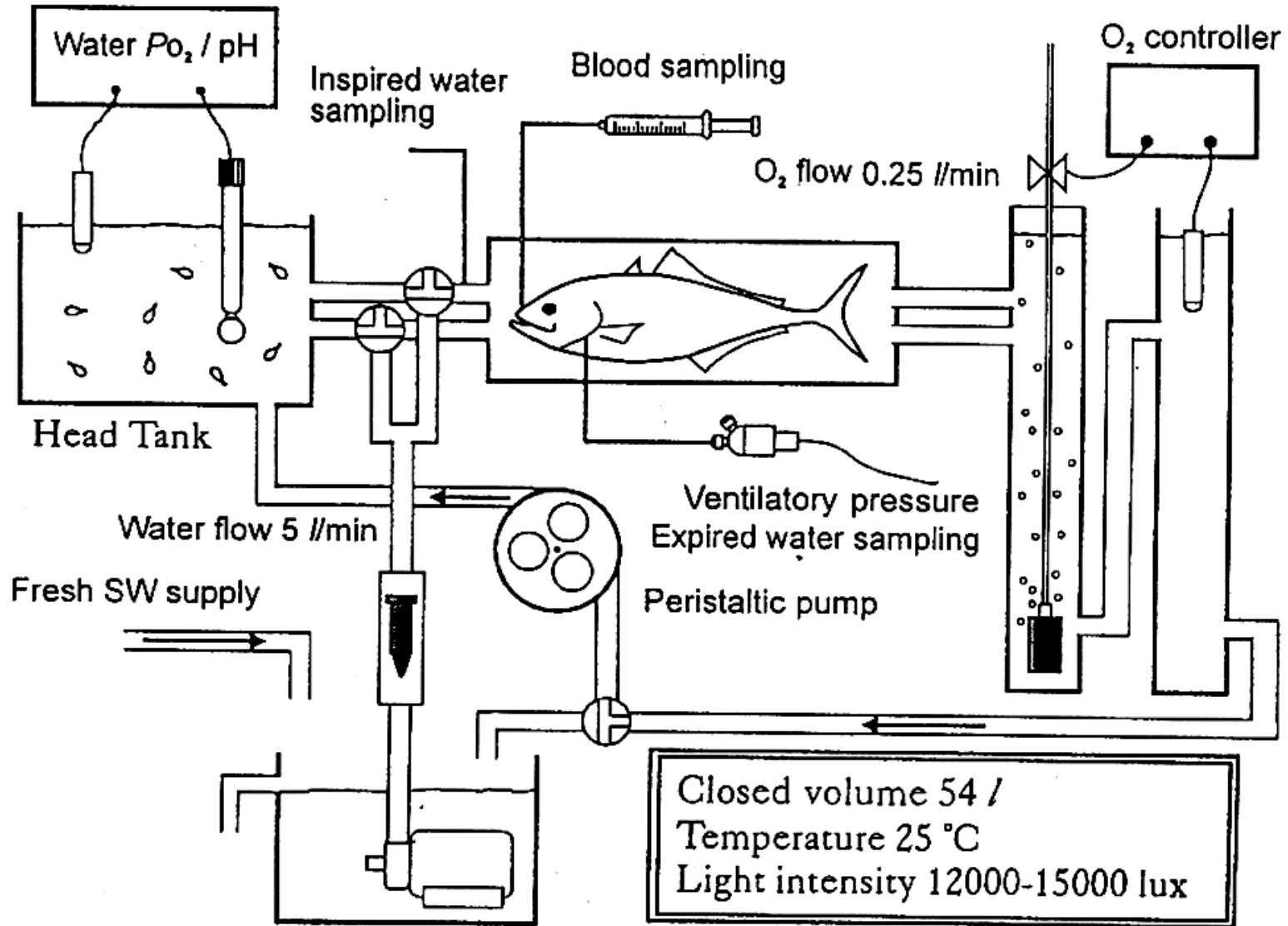
Indirect immunofluorescence staining of *C. marina* with anti-glycocalyx antiserum and FITC-labeled secondary antibody

Phase-contrast

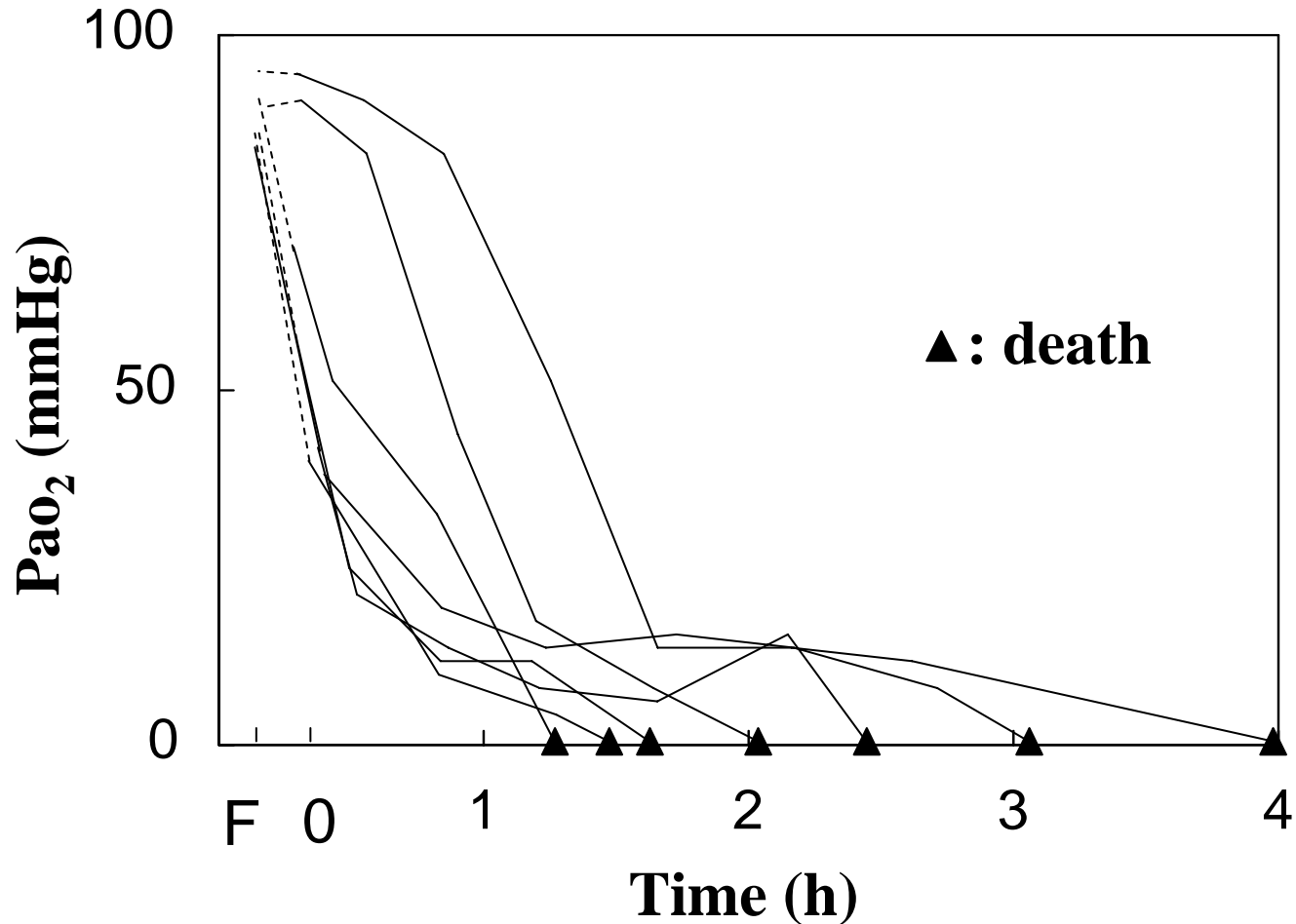
Fluorescence



The system for monitoring blood oxygen pressure of yellowtail after exposure to *C. marina* (from Dr. Ishimatsu)



Changes in oxygen partial pressure of arterial blood (P_{aO_2}) in yellowtail exposed to *C. marina* (from Dr. Ishimatsu)

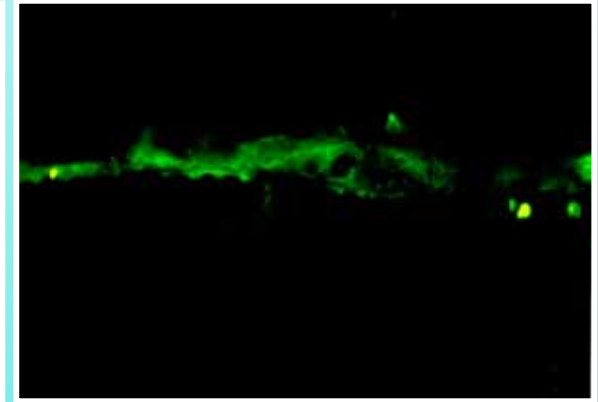


Indirect immunofluorescence staining of gill tissues of control and *C. marina*-exposed yellowtails with anti-glycocalyx antiserum and FITC-labeled secondary antibody

Control

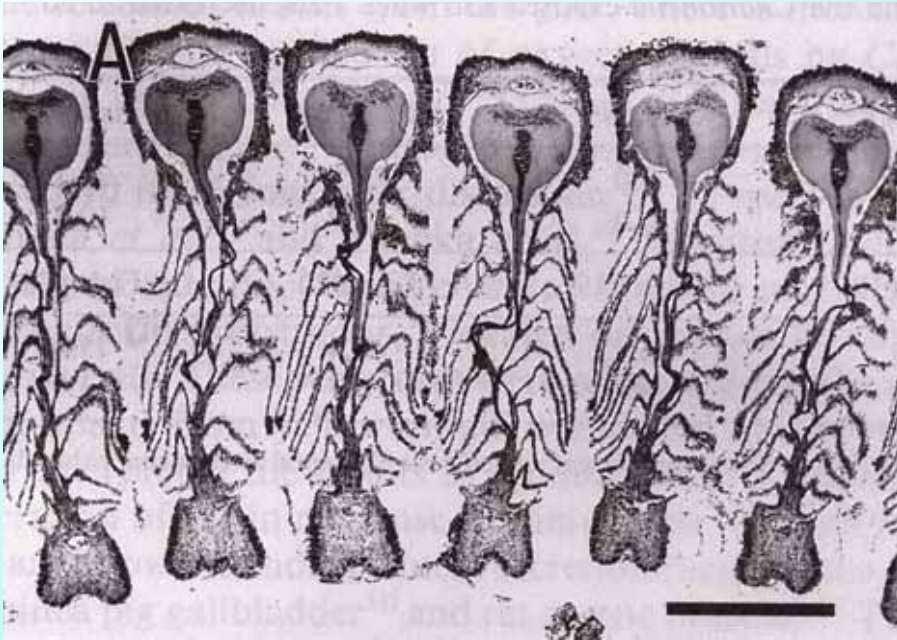


***C. marina*-exposed**

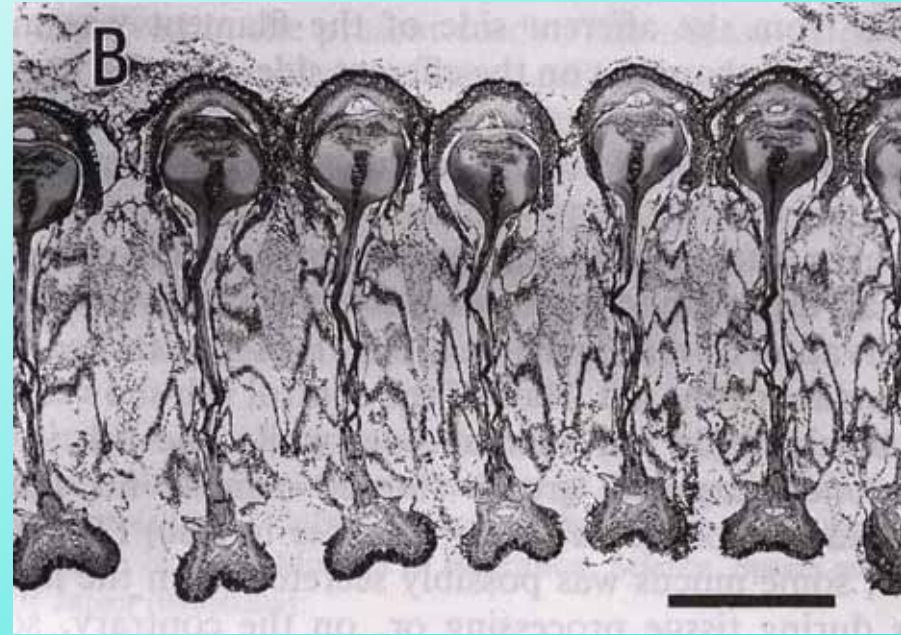


Gill tissue damage after exposure to *C. marina*

Normal gill



Gill after exposure to *C. marina*

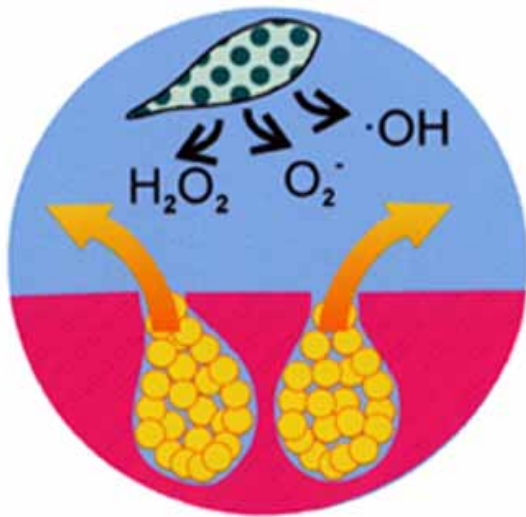


Provided by Dr. Ishimatsu

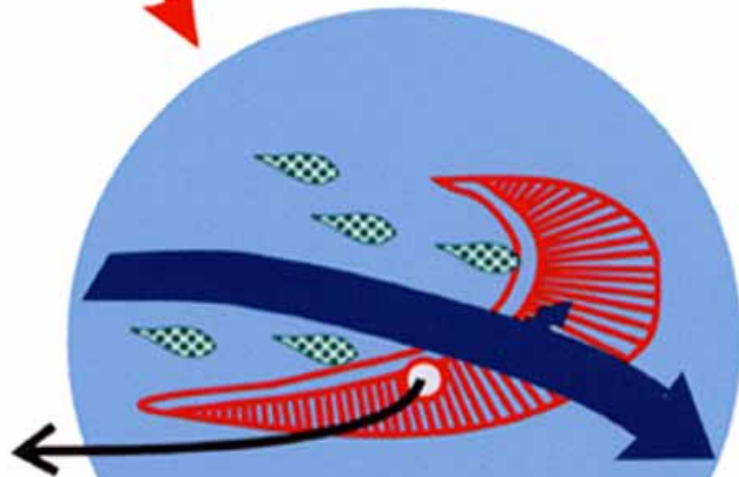
The earliest physiological changes observed in yellowtail after exposure to *C. marina* is the decrease in arterial oxygen partial pressure.

The primary target of *C. marina* is gill.

Chattonella



Gill epithelium



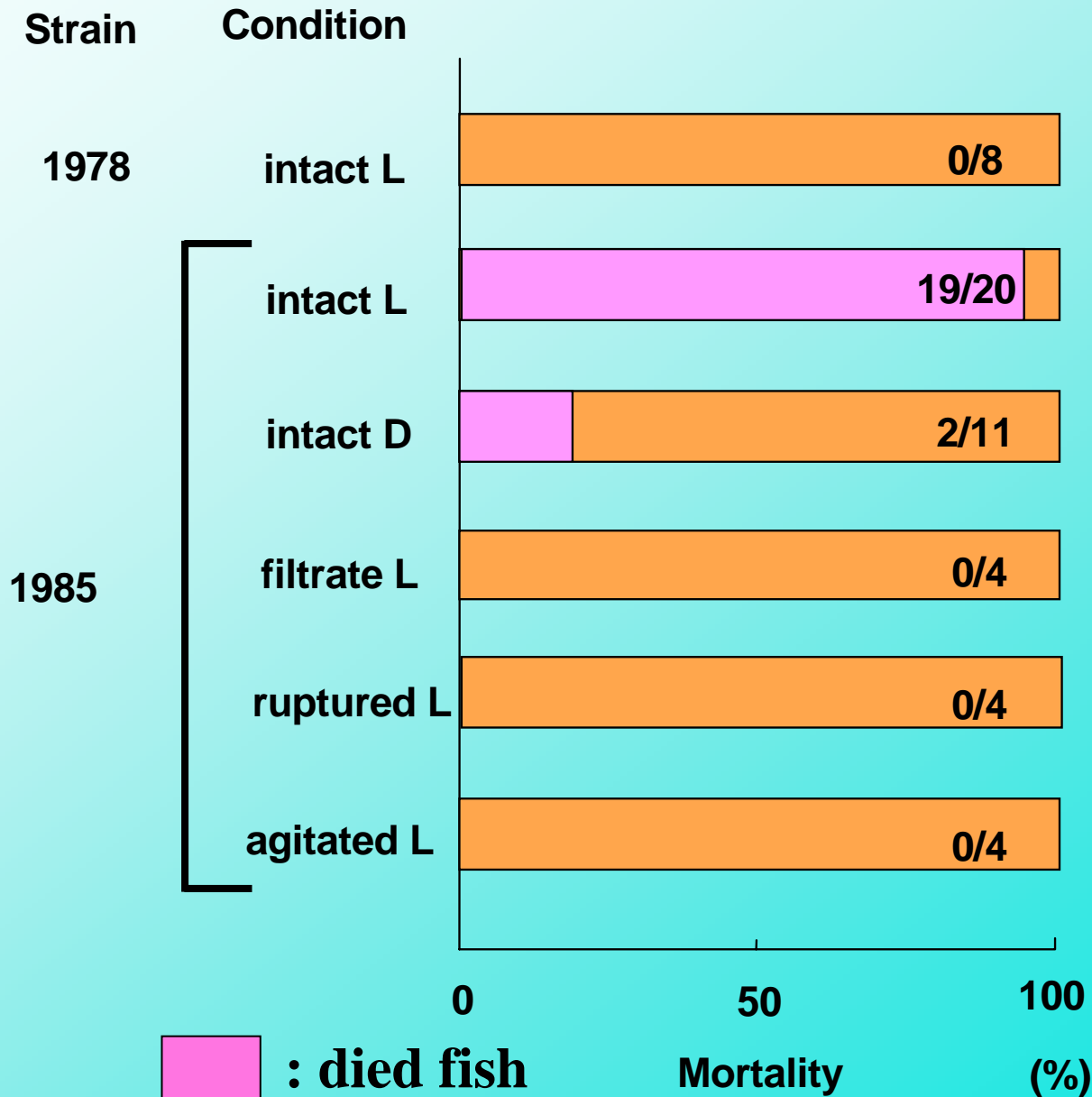
**Respiratory water shunt
+ Other disturbances?**

**What is the evidence for
the involvement of
ROS in gill tissue
damage and eventual
fish-killing by *C.
marina*?**

Fish (yellowtail) mortality within 8 h of exposure to *C. marina*.

Strain 1978 : low ROS generation

Strain 1985 : High ROS generation



Processes leading to fish death by *C. marina*

ROS-mediated gill tissue damage (gill surface might be covered with glycocalyx discharged from *C. marina*)



Continuous ROS generation by glycocalyx might induce over secretion of mucus substances on gill tissue



Dysfunction of gill



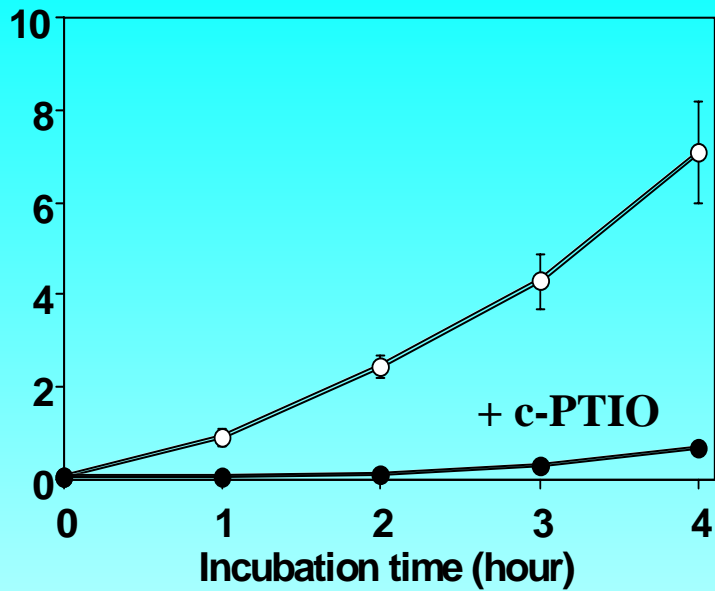
Decrease in the arterial oxygen partial pressure



Suffocation

**Are there any other
possible toxic
factors?**

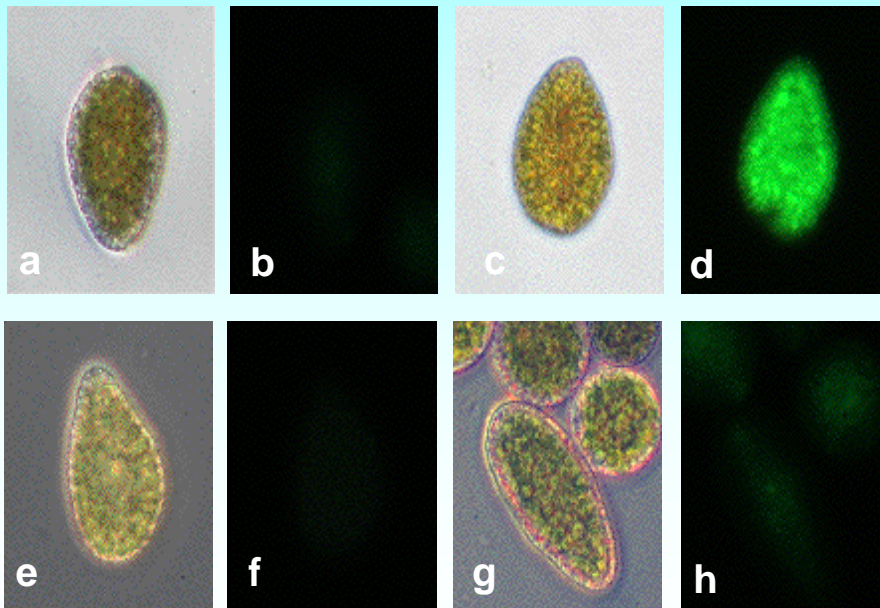
**In addition to ROS
such as H_2O_2 , O_2^- , and
 $\bullet\text{OH}$ radical, *C. marina*
produces **nitric oxide**
(NO)**



(A)

Nitric oxide (NO) production by *C. marina*. (A) Kinetics of NO production detected by NO-specific fluorescence probe (DAF-FM DA).

(B)



None

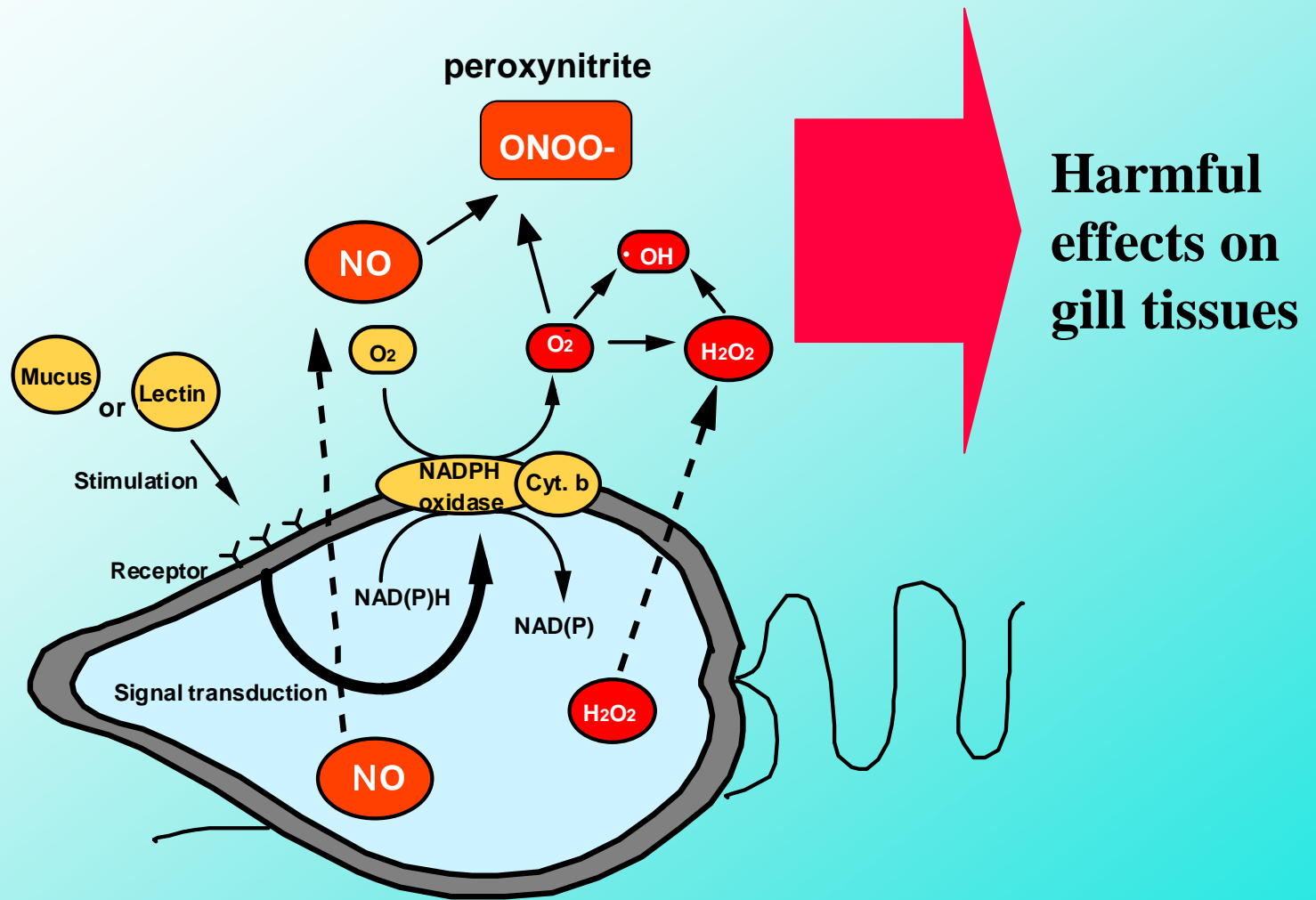
+ c-PTIO

Fluorescence observation of *C. marina* in the presence or absence of specific NO scavenger, carboxy-PTIO

0 time

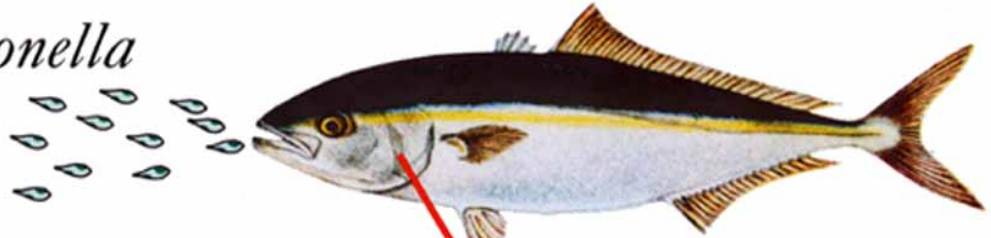
After 3 h

Generation of reactive oxygen species in *Chattonella marina*



ROS-mediated toxic mechanism of *C. marina*; possible involvement of NO

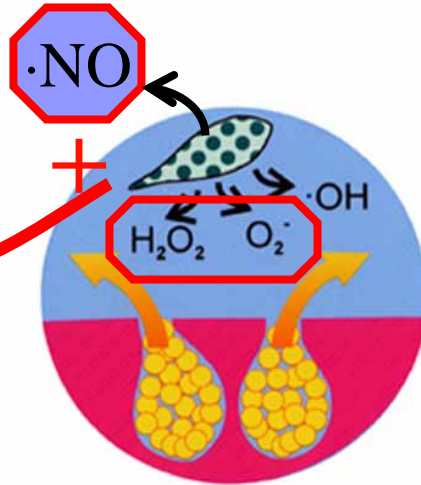
Chattonella



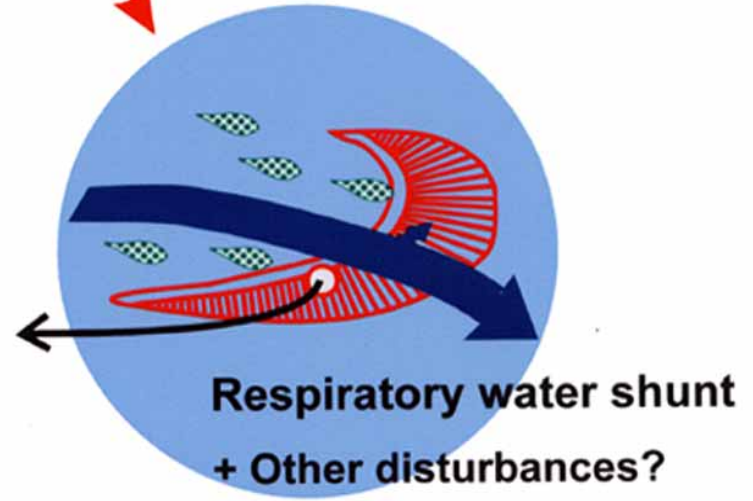
Hemoglobin (Hb)

Peroxynitrite (ONOO⁻)

ONOO⁻



Gill epithelium



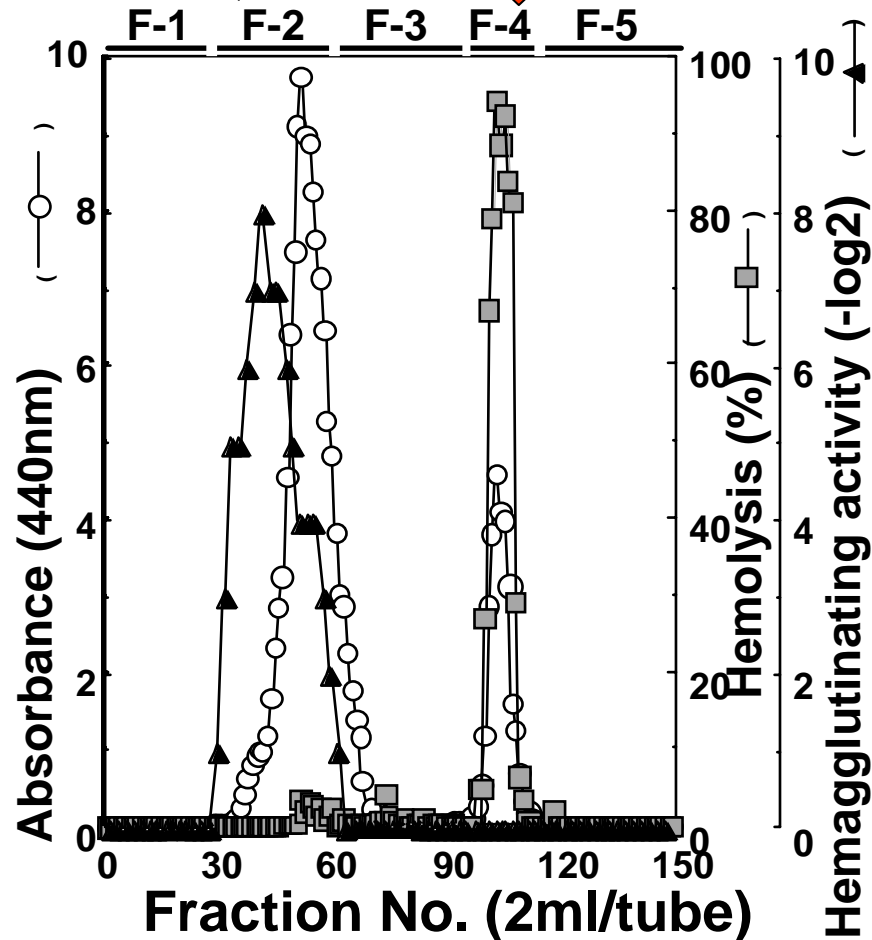
Methemoglobin (MetHb)

Cannot bind oxygen !!

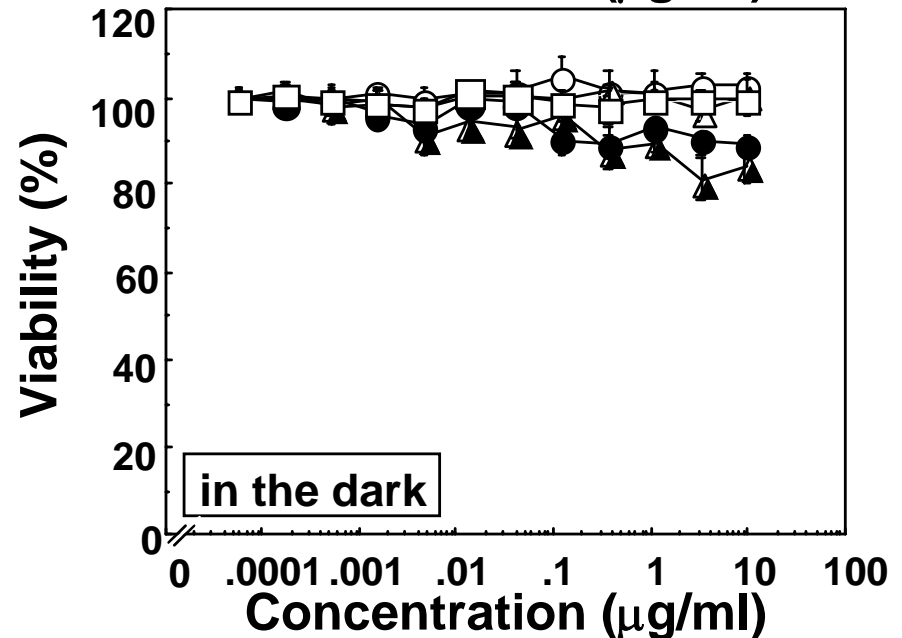
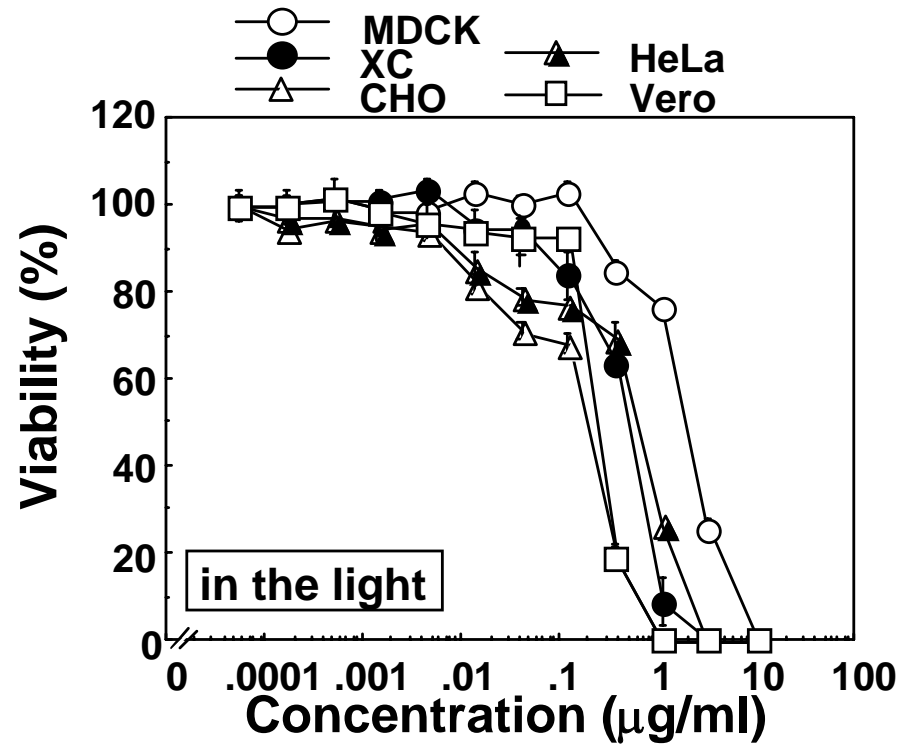
Hemolytic toxins in *C. marina*

Hemolytic and hemagglutinating activities in the methanol extract of *C. marina*

Hemagglutinin ↓ Hemolytic toxin ↓

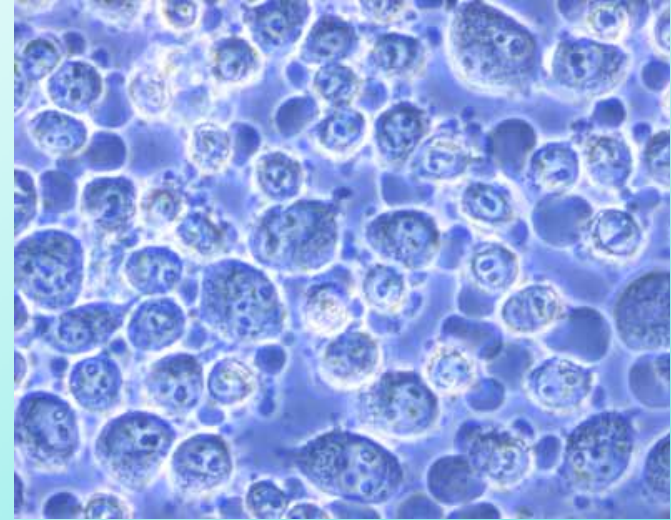
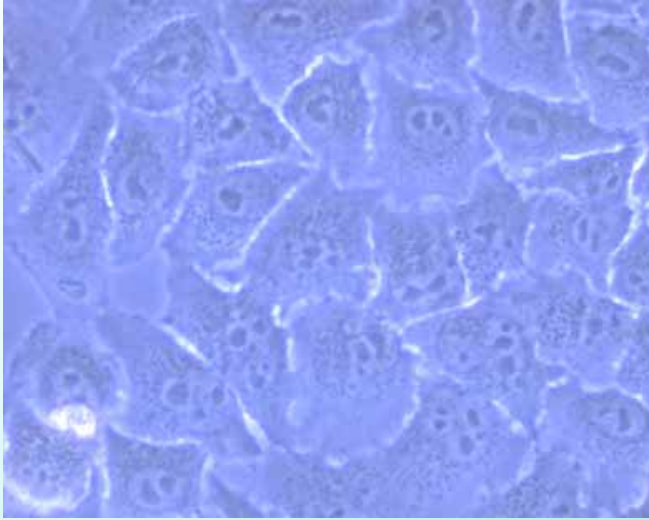


**Hemolytic
toxin from *C.
marina* shows
light-
dependent
cytotoxicity on
various cell
lines**

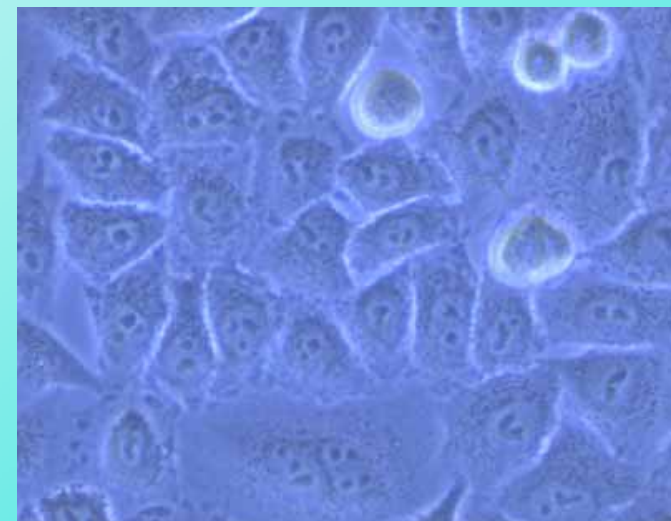
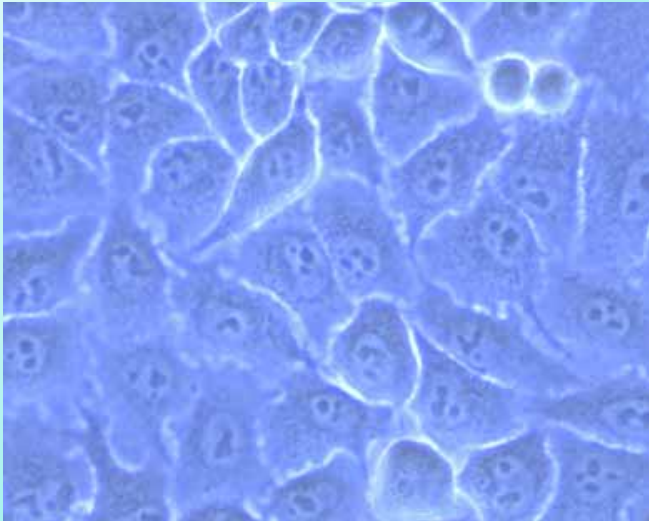


Homolytic toxin induced light-dependent morphological changes in HeLa cells

light



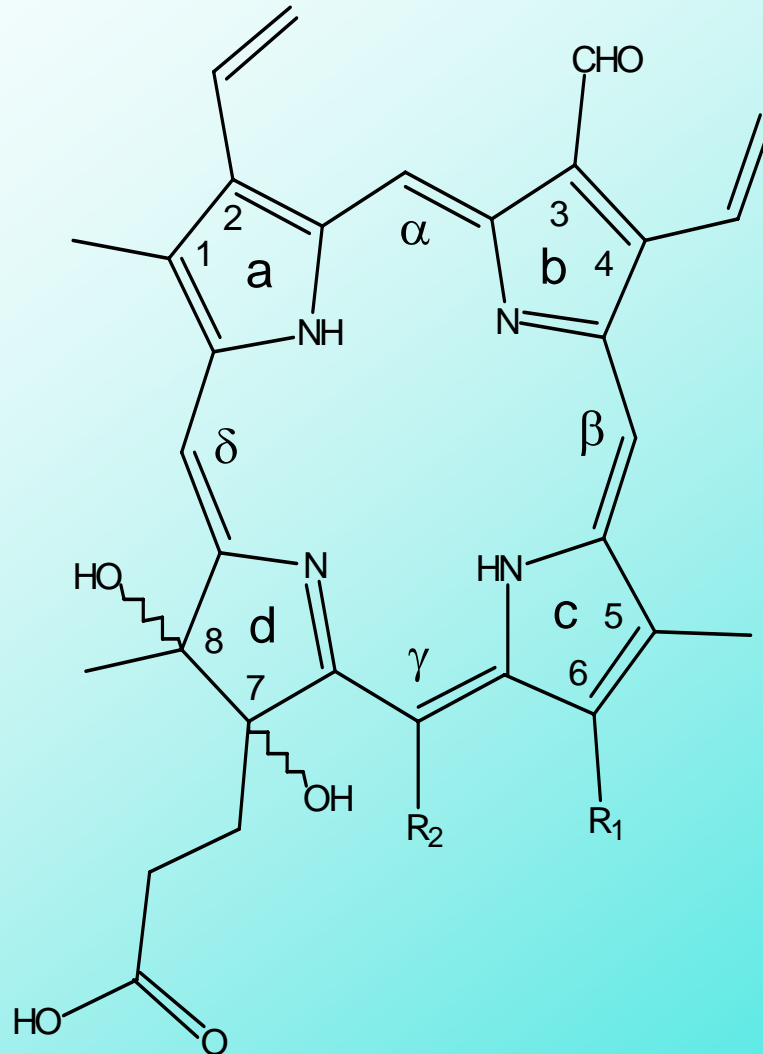
dark



0 time

30 min

Light-dependent hemolytic toxin isolated from *C. marina* may be a porphyrin derivative

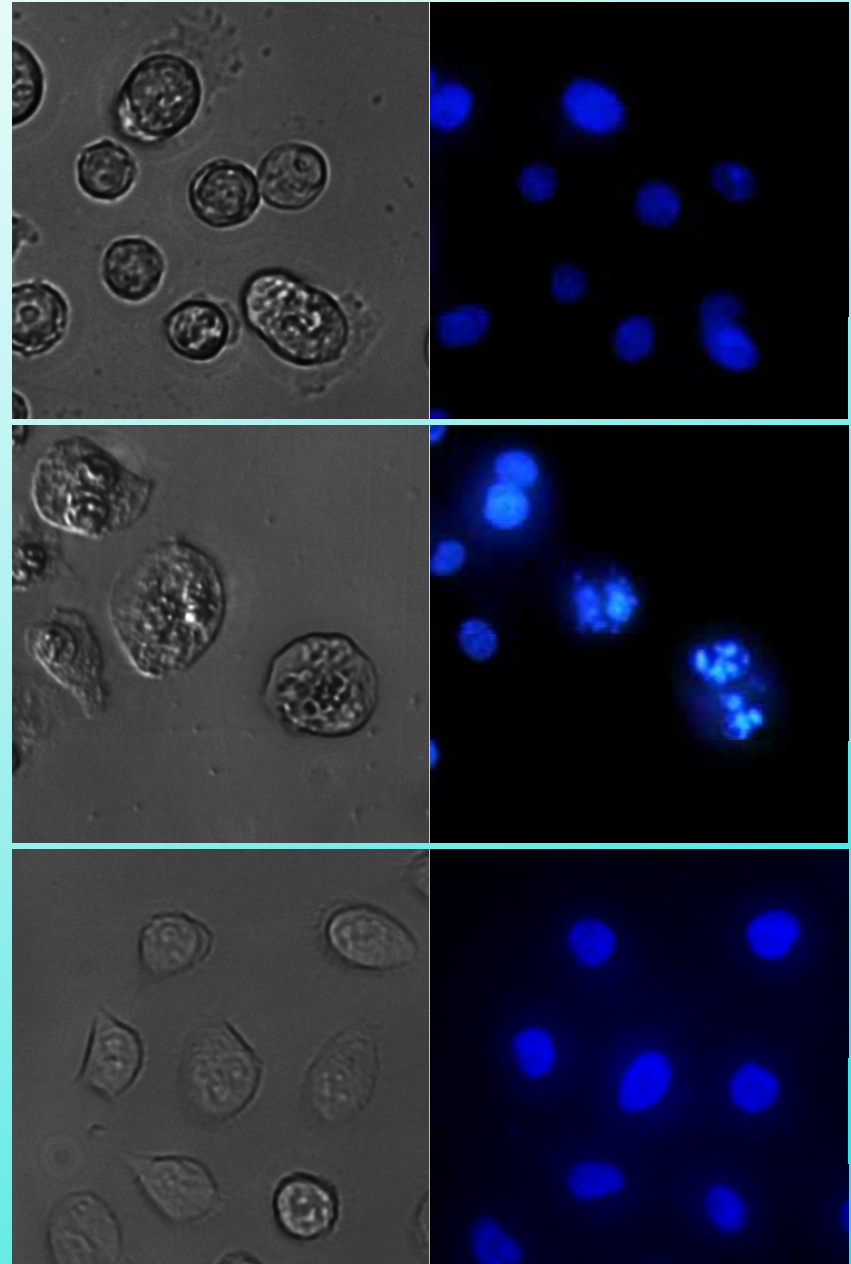


Hemolytic
toxin
isolated
from *C.*
marina
induces
necrosis
in HeLa
cells

+
hemolytic
toxin
from *C.*
marina

+
pheophorbide

control



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